



Product Sheet

M17/5.2 (ATCC® TIB-237™)

Please read this FIRST



Storage Temp.
liquid nitrogen
vapor phase



Biosafety Level
1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

The base medium for this cell line is ATCC-formulated Dulbecco's Modified Eagle's Medium, Catalog No. 30-2002. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: M17/5.2 (ATCC® TIB-237™)

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Or contact your local distributor

Description

Organism: *Rattus norvegicus* (B cell); *Mus musculus* (myeloma), rat (B cell); mouse (myeloma)

Isotype: rat IgG2b kappa

Tissue: spleen

Cell Type: hybridoma: B lymphocyte

Morphology: lymphoblast

Growth Properties: suspension

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C , preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

1. Initiate culture as soon as possible upon receipt.
2. Thaw by rapid agitation in 37°C water bath. Thawing should be rapid (within 40-60 seconds). As soon as the ice is melted, remove the ampule from the water bath and immerse in 70% ethanol at room temperature. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the cell suspension and dilute it with the recommended culture medium in a culture flask (see specific batch information above for dilution ratio); incubate at 37°C with 10% CO_2 in air atmosphere. Since it is important to avoid excessive alkalinity of the medium during recovery of the cells, it is suggested that the culture medium be placed into the culture flask, tube, etc. and the pH be adjusted, as necessary, prior to the addition of the ampule contents. Note that the bicarbonate content of the culture medium will determine whether an atmosphere containing CO_2 will be required.
4. It is not necessary to remove the freezing additive. However, if desired, the culture medium may be changed to remove the protective freezing additive (dimethylsulfoxide) 24 hours after thawing. If it is desired that the freezing additive be removed immediately, or that a more concentrated cell suspension be obtained, centrifuge the above diluted suspension at approximately $125 \times g$ for 10 minutes, discard the fluid and resuspend the cells with growth medium at the dilution ratio given in the specific batch information above.

Handling Procedure for Flask Cultures

The flask was seeded with cells (see specific batch information above for concentration), grown and completely filled with medium to prevent loss of cells in transit.

1. Upon receipt incubate the flask in an upright position for several hours to return the flask contents to 37°C .
2. After the temperature has equilibrated, aseptically remove the entire contents of the flask and centrifuge at $300 \times g$ for 15 minutes.
3. Resuspend the cell pellet in 10-12 mL of the shipping medium. From this suspension remove a sample for a cell count and viability so that the cell density of the suspension can be adjusted to $2-3 \times 10^5$ viable cells/mL. If the suspension needs to be diluted use the shipping medium.
4. Incubate the culture in a flat position at 37°C . The shipping medium contains reduced sodium bicarbonate suitable for a 5% CO_2 in air incubator. DMEM usually contains 3.7 grams of sodium bicarbonate per liter and should be incubated in a 10% CO_2 in air incubator.
5. Maintain the cell density of the culture as suggested under the subculture procedure described above.

Subculturing Procedure



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Cultures can be maintained by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 1×10^5 viable cells/mL.

Interval: Maintain between 1×10^5 and 1×10^6 cells/mL.

Medium Renewal: Every 2 to 3 days



Cryopreservation Medium

Complete growth medium supplemented with 5% (v/v) DMSO.



Comments

Animals were immunized with mouse cytotoxic T lymphocytes (CTL).

Spleen cells were fused with NS-1 myeloma cells.

The antibody blocks CTL mediated cytotoxicity, and can be used to precipitate LFA-1 from lysates of CTLs.



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Additional information on this culture is available on the ATCC web site at www.atcc.org.

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