



Product Sheet

Pseudoharpagon pertyi (ATCC® PRA-359™)

Please read this **FIRST**

Storage Temp.
Frozen Cultures:
-70°C for 1 week;
liquid N₂ vapor
for long term
storage



Freeze-dried
Cultures:
2-8°C

Live Cultures:
See Protocols
section for
handling
information



Biosafety Level
1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Pseudoharpagon pertyi* (ATCC® PRA-359™)

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Or contact your local distributor



Description

Strain Designation: NY0199
Depositor: N Yubuki
Isolation: Beach sand, Bamfield, Canada, June 2009



Propagation

Growth Conditions

Temperature: 25°C

Culture System : Xenic, grown with mixed bacteria in the dark. Dilute ATCC medium 1171 1:20 in artificial seawater.

Medium

ATCC® Medium 1171: TYGM-9 medium

Instructions for Complete Medium

Media: ATCC Medium 1171 diluted to 5% strength in artificial seawater (ASW)



Protocols

Storage and Culture Initiation

Frozen ampules packed in dry ice should either be thawed immediately or stored in liquid nitrogen. If liquid nitrogen storage facilities are not available, frozen ampules may be stored at or below -70°C for approximately one week. **Do not under any circumstance store frozen ampules at refrigerator freezer temperatures (generally -20°C).** Storage of frozen material at this temperature will result in the death of the culture.

1. To thaw a frozen ampule, place it in a 35°C water bath such that the lip of the ampule remains above the water line. Thawing time is approximately 2 to 3 minutes. Do not agitate the ampule. Do not leave ampule in water bath after it is thawed.
2. Immediately after thawing, aseptically transfer contents to a 16 x 125 mm screw-capped test tube containing 6-8 ml of ATCC Medium 1171 diluted to 5% strength in artificial seawater (ASW).
3. Screw the cap on tightly and incubate on a 15° horizontal slant at 25°C.

Culture Maintenance

1. Prepare 5% diluted ATCC Medium 1171 as indicated above.
2. When the culture is at or near peak density, rub the surface of the tube with a sterile cotton swab, and agitate the swab to dislodge the adherent cells. Invert gently 10 times to distribute cells evenly.
3. Transfer approximately 0.25 ml to a 16 x 125 mm screw-capped test tube containing 6-8 ml of fresh ATCC Medium 1171 diluted to 5% strength in artificial seawater (ASW).
4. Screw the cap on tightly and incubate on a 15° horizontal slant at 25°C.



Cryopreservation

Harvest and Preservation

1. Harvest the cells from a culture that is at or near peak density by centrifuging at 900 x g for 5 minutes.
2. If the cell concentration exceeds the required level do not centrifuge, but adjust the concentration to between 2 x 10⁶ and 2 x 10⁷ cells/ml with fresh medium. If the concentration is too low, centrifuge at 900 x g for 5 min and resuspend the pellet in the volume of fresh medium required to yield the desired concentration.
3. While cells are centrifuging prepare a 15% (v/v) solution of sterile DMSO as follows: Add the required volume of DMSO to a glass screw-capped test tube and place it in an ice bath. Allow the DMSO to solidify. Add the required volume of refrigerated medium. Dissolve the DMSO by inverting the tube several times.
NOTE: If the DMSO solution is not prepared on ice, an exothermic reaction will occur that may precipitate certain components of the medium.
4. Mix the cell preparation and the DMSO in equal portions. Thus, the final concentration will be between 10⁶ and 10⁷ cells/ml and 7.5% (v/v) DMSO. The time from the mixing of the cell preparation and DMSO stock solution to the start of the freezing process should be no less than 15 min and no longer than 30 min.
5. Dispense in 0.5 ml aliquots into 1.0 - 2.0 ml sterile plastic screw-capped cryules (special plastic vials for cryopreservation).



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- Place vials in a controlled rate freezing unit. From room temperature cool at -1°C/min to -40°C. If freezing unit can compensate for the heat of fusion, maintain rate at -1°C/min through heat of fusion. At -40°C plunge ampules into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately -1°C/min.)
- The frozen preparations are stored in either the vapor or liquid phase of a nitrogen freezer.
- To establish a culture from the frozen state place an ampule in a water bath set at 35°C (2-3 min). Immerse the vial just sufficiently to cover the frozen material. Do not agitate the vial.
- Immediately after thawing, aseptically transfer contents to a 16 x 125 mm screw-capped test tube containing 6-8 ml of ATCC Medium 1171 diluted to 5% strength in artificial seawater (ASW).
- Screw the cap on tightly and incubate on a 15° horizontal slant at 25°C.
- Follow the protocol for maintenance of culture.



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

ATCC Warranty

The viability of ATCC® products is warranted for 30 days from the date of shipment, and is valid only if the product is stored and cultured according to the information included on this product information sheet. ATCC lists the media formulation that has been found to be effective for this strain. While other, unspecified media may also produce satisfactory results, a change in media or the absence of an additive from the ATCC recommended media may affect recovery, growth and/or function of this strain. If an alternative medium formulation is used, the ATCC warranty for viability is no longer valid.

Disclaimers

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.

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