**Strain Designation:** PW19 (P19)  
**Deposited Name:** Oblongichytrium sp.  
**Depositor:** M Berbee  
**Isolation:** Marine invertebrate gut contents  
**Isolated by:** W Marshall, 2004  
**Bamfield, British Columbia, Canada**

### Growth Conditions
- **Growth condition:** axenic  
- **Temperature:** 15.0°C  
- **Medium:** ATCC® Medium 2673: Thraustochytrid medium

### Instructions for Complete Medium
ATCC Medium 2673

### Culture Maintenance
1. When the culture is at or near peak density, vigorously agitate the culture.
2. Transfer approximately 0.25 ml to a fresh tube or flask containing 5 ml of fresh ATCC medium 2673.
3. Screw the caps on tightly and incubate at 15°C (incubate test tubes at a 15° horizontal slant).
4. Thraustochytrids will eventually form an almost continuous sheet of cells on the bottom surface of the flask or test tube, with other cells floating in the liquid column and collecting at the fluid meniscus.
5. Repeat steps 1-3 at 2-3 week intervals.

1. To achieve the best results set up cultures with several different inocula (e.g. 0.25 ml, 0.5 ml, 1.0 ml). Harvest cultures and pool when the culture that received the lowest inoculum is at or near peak density.
2. Transfer approximately 0.25 ml to a fresh tube or flask containing 5 ml of fresh ATCC medium 2673.
3. Screw the caps on tightly and incubate at 15°C (incubate test tubes at a 15° horizontal slant).
4. Thraustochytrids will eventually form an almost continuous sheet of cells on the bottom surface of the flask or test tube, with other cells floating in the liquid column and collecting at the fluid meniscus.
5. Repeat steps 1-3 at 2-3 week intervals.

### Cryopreservation
1. To achieve the best results set up cultures with several different inocula (e.g. 0.25 ml, 0.5 ml, 1.0 ml). Harvest cultures and pool when the culture that received the lowest inoculum is at or near peak density.
2. If the cell concentration exceeds the required level do not centrifuge, but adjust the concentration to between 2 x 10^6 and 2 x 10^7 cells/ml with fresh medium. If the concentration is too low, centrifuge at 600 x g for 5 min and resuspend the pellet in the volume of fresh medium required to yield the desired concentration.
3. While cells are centrifuging prepare a 20% (v/v) solution of sterile DMSO as follows: Add the required volume of DMSO to a glass screw-capped test tube and place it in an ice bath. Allow the DMSO to solidify. Add the required volume of refrigerated medium. Dissolve the DMSO by inverting the tube several times.
4. Mix the cell preparation and the DMSO in equal portions. Thus, the final concentration will be between 10^6 and 10^7 cells/ml and 10% (v/v) DMSO. The time from the mixing of the cell preparation and DMSO stock solution to the start of the freezing process should be no less than 15 min and no longer than 30 min.
5. Dispense in 0.5 ml aliquots into 1.0 - 2.0 ml sterile plastic screw-capped cryules (special plastic vials for cryopreservation).
6. Place the vials in a controlled rate freezing unit. From room temperature cool at -1°C/min to -40°C. If the freezing unit can compensate for the heat of fusion, maintain rate at -1°C/min through the heat of fusion. At -40°C plunge into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately -1°C/min.)
7. The frozen preparations are stored in either the vapor or liquid phase of a nitrogen freezer.
8. To establish a culture from the frozen state place an ampule in a water bath set at 35°C (2-3 min). Immerse the vial just sufficient to cover the frozen material. Do not agitate the vial.
9. Immediately after thawing, aseptically remove the contents of the ampule and inoculate into 5 ml of fresh ATCC medium 2673 in a 16 x 125 mm screw-capped test tube. Incubate at 15°C.

### References
 References and other information relating to this product are available online at www.atcc.org.

**Biosafety Level: 1**

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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