



## Product Sheet

# SW 872 [SW-872, SW872] (ATCC® HTB-92™)

### Please read this FIRST



Storage Temp.  
**liquid nitrogen**  
vapor phase

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Biosafety Level  
**1**

### Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

### Complete Growth Medium

The base medium for this cell line is ATCC-formulated Leibovitz's L-15 Medium, Catalog No. 30-2008. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.

(Note: The L-15 medium formulation was devised for use in a free gas exchange with atmospheric air. A CO<sub>2</sub> and air mixture is detrimental to cells when using this medium for cultivation)

### Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: SW 872 [SW-872, SW872] (ATCC® HTB-92™)

American Type Culture Collection  
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Manassas, VA 20108 USA  
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## Description

**Organism:** *Homo sapiens*, human

**Tissue:**

connective tissue

**Disease:** liposarcoma

**Age:** 36 years adult

**Gender:** male

**Morphology:** fibroblast

**Growth Properties:** adherent

**Isoenzymes:**

AK-1, 1

ES-D, 1

G6PD, B

GLO-I, 1-2

PGM1, 1-2

PGM3, 1

**Cytogenetic Analysis:** hypertriploid; modal number = 80; range = 66 to 81. The rate of higher ploidies was 8.2%. Ten markers were common to most cells. These were: der(5)t(5;?)(q31;?)1, der(5)t(5;?)(q31;?)2, der(6)t(6;?)(q15;?), der(7)t(7;?)(q36;?), t(15q16q) and five others. Both der(5) markers, the der(7) and t(15q16q) were paired. There were 5 copies of N20 and N21, 4 copies of N8, N9, N11, N14 and N17 and a single copy of X in each cell.

## Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

## SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

## Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

## Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. It is recommended that the cryoprotective agent be removed immediately. Centrifuge the cell suspension at approximately 125 xg for 5 to 10 minutes. Discard the supernatant and resuspend the cell pellet in an appropriate amount of fresh growth medium.
3. Transfer the cells to an appropriate size vessel.
4. Incubate the culture at 37°C in a suitable incubator. The L-15 medium formulation was devised for use in a free gas exchange with atmospheric air. A CO<sub>2</sub> and air mixture is detrimental to cells when using this medium for cultivation.

## Subculturing Procedure

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin- 0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).

Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.



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4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cultures at 37°C.

**Subcultivation Ratio:** A subcultivation ratio of 1:3 to 1:6 is recommended

**Medium Renewal:** 2 to 3 times per week



### Cryopreservation Medium

Complete growth medium described above supplemented with 5% (v/v) DMSO.  
Cell culture tested DMSO is available as ATCC Catalog No. 4-X.



### Comments

The histopathology evaluation reported an undifferentiated malignant tumor consistent with liposarcoma. An ampule at passage 4 was received at the ATCC in January, 1982.



### References

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).



### Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).

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