



Product Sheet

NCI-H82 [H82] (ATCC® HTB-175™)

Please read this FIRST



Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

The base medium for this cell line is ATCC-formulated RPMI-1640 Medium, ATCC [30-2001](#). To make the complete growth medium, add the following components to the base medium: fetal bovine serum (ATCC [30-2020](#)) to a final concentration of 10%.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: NCI-H82 [H82] (ATCC® HTB-175™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor

Description

Organism: *Homo sapiens*, human

Tissue: lung; derived from metastatic site: pleural effusion

Disease: carcinoma; small cell lung cancer

Age: 40 years

Gender: male

Morphology: epithelial

Growth Properties: aggregates in suspension; the cells grow in very large aggregates, and the aggregates are the only viable cell population

Isoenzymes:

AK-1, 1

ES-D, 1

G6PD, B

GLO-I, 1

Me-2, 1

PGM1, 1-2

PGM3, 1-2

DNA Profile:

Amelogenin: X

CSF1PO: 11

D13S317: 8

D16S539: 12

D5S818: 12

D7S820: 10,13

THO1: 9,9.3

TPOX: 11

vWA: 14

Cytogenetic Analysis: This is a near triploid human cell line. The modal chromosome number is 58, occurring at 44% with polyploidy at 3%. Marker chromosomes der(1)t(1;709p13;p11), t(13q;?HSR;15q) and der(190t(19;?)(q13.4;?) were common to most cells. There were two distinct subpopulations readily distinguished by karyotype. Besides uniform changes in the numbers of copies of some normal chromosomes, one population had der(3)t(3;20)(p11;p11?), t(3q19p), i(7q) and a minute chromosome of unknown origin. The other had t(1q17p), del(1)(q21), der(3)t(3;7)(p12;q11) plus two other markers. Each cell had two copies of a normal X chromosome. The Y chromosome was not detected in Q banded preparations.

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium. and spin at



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approximately 125 xg for 5 to 7 minutes.

- Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio), and dispense into a 25 cm² or a 75 cm² culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
- Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.



Handling Procedure for Flask Cultures

The flask was seeded with cells (see specific batch information), grown, and completely filled with medium at ATCC to prevent loss of cells during shipping.

- Upon receipt visually examine the culture for macroscopic evidence of any microbial contamination. Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination
- Incubate the flask in an upright position for several hours at 37°C. After the temperature has equilibrated, aseptically remove the entire contents of the flask and centrifuge at 125 xg for 5 to 10 minutes. Remove shipping medium and save for reuse. Resuspend the cell pellet in 10 mL of this medium.
- From this cell suspension remove a sample for a cell count and viability. Adjust the cell density of the suspension to 2-5 x 10⁵ viable cells/mL in the shipping medium.
- Incubate the culture, horizontally, at 37°C in a 5% CO₂ in air atmosphere. Maintain the cell density of the culture as suggested under the subculture procedure.



Subculturing Procedure

This line grows as aggregates of cells in suspension. Culture can be maintained by addition of medium or by replacement of medium. Alternatively, the cells may be collected by centrifugation and dispersed into fresh medium.

Subcultivation Ratio: A subcultivation ratio of 1:2 to 1:5 is recommended

Medium Renewal: 2 to 3 times per week



Cryopreservation Medium

Complete culture medium described above supplemented with 5% (v/v) DMSO.

Cell culture tested DMSO is available as ATCC Catalog No. 4-X.



Comments

The morphology of the original tumor was not characteristic of SCLC.

The line is a biochemical and morphological variant of SCLC that expresses neuron specific enolase and the brain isoenzyme of creatine kinase.

It does not have detectable levels of L-DOPA decarboxylase or bombesin.

The cells produce an abnormally sized p53 mRNA (3.7 kb).

C-myc DNA sequences are amplified about 25 fold, and there is a 24 fold increase in c-myc RNA relative to normal cells.

The cells are reported to express functional ANP receptors, but treatment with ANP does not alter their growth pattern.

The cells stain positively for neurofilaments and vimentin.

There is expression of v-fes, v-fms, Ha-ras, Ki-ras, N-ras and c-raf 1 mRNAs.



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

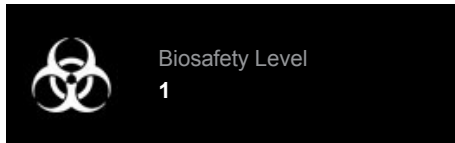
ATCC Warranty



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Biosafety Level
1

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Disclaimers

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Additional information on this culture is available on the ATCC web site at www.atcc.org.
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