



Product Sheet

MDA-MB-415 (ATCC® HTB-128™)

Please read this FIRST



Storage Temp.
liquid nitrogen
vapor phase



Biosafety Level
1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

Leibovitz's L-15 medium with 2mM L-glutamine supplemented with 10 mcg/ml insulin and 10 mcg/ml glutathione, 85%; fetal bovine serum, 15%. (Note: The L-15 medium formulation was devised for use in a free gas exchange with atmospheric air. A CO₂ and air mixture is detrimental to cells when using this medium for cultivation)

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: MDA-MB-415 (ATCC® HTB-128™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor

Description

Organism: *Homo sapiens*, human

Tissue: mammary gland/breast; derived from metastatic site: pleural effusion

Disease: adenocarcinoma

Age: 38 years

Gender: female

Morphology: epithelial

Growth Properties: adherent

Isoenzymes:

AK-1, 1

ES-D, 2

G6PD, A

GLO-I, 1-2

Me-2, 2

PGM1, 1

PGM3, 1

DNA Profile:

Amelogenin: X

CSF1PO: 10,12

D13S317: 11,13

D16S539: 13

D5S818: 11,13

D7S820: 9,10

THO1: 7,9.3

TPOX: 8

vWA: 17

Cytogenetic Analysis: modal number =73; range = 65 to 76. The stemline chromosome number is hypertriploid with the 2S component occurring at 0.8%. Ten markers (del(2), t(6;?), t(6;?), t(2;12), t(14;?), and single or paired M6, M7, M8, M9, and M10) were common to all cells, and four others (M13, M15, M16, and t(13,21) occurred in some cells. The X chromosome was generally tetrasomic and chromosome 4 was tetrasomic or pentasomic (P35). No Y chromosomes were detected in QM stained preparations.

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.


1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio) and dispense into a 25 cm² or a 75 cm² culture flask.




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5. Incubate the culture at 37°C in a suitable incubator. The L-15 medium formulation was devised for use in a **free gas exchange with atmospheric air**. A CO₂ and air mixture is detrimental to cells when using this medium for cultivation



Subculturing Procedure

Subcultures are prepared by scraping. Remove old medium, add fresh, dislodge cells, aspirate and dispense into new flasks. Subculture every 6 to 8 days.

Subcultivation Ratio: A subcultivation ratio of 1:2 is recommended

Medium Renewal: 2 to 3 times per week



Cryopreservation Medium

Complete growth medium described above supplemented with 5% (v/v) DMSO.
Cell culture tested DMSO is available as ATCC Catalog No. 4-X.



Comments

The cells express the WNT7B oncogene [PubMed: 8168088].

The patient presenting with the tumor was from Paraguay and, although listed as Caucasian, may have had mixed ancestry as suggested by the presence of G6PD type A phenotype in the cells.

The line forms flat, spreading plaques of epithelia which exhibit desmosomes, extensive microtubules and microfilaments upon examination with the electron microscope.

It is not amenable to trypsinization.



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.

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