



Product Sheet

MECA-79 (ATCC® HB-9479™)

Please read this FIRST



Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

RPMI 1640 medium with 2 mM L-glutamine adjusted to contain

1.5 g/L sodium bicarbonate, 4.5 g/L glucose, 10 mM HEPES, and 1.0 mM sodium

pyruvate, 90%; fetal bovine serum, 10%; supplemented with 0.05 mM

2-mercaptoethanol. Note: A feeder layer of mouse peritoneal exudate or

irradiated murine fibroblasts (e.g. irradiated STO cells, ATCC catalog # X-56)

is required for recovery and growth of the MECA-79.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: MECA-79 (ATCC® HB-9479™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
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Or contact your local distributor

Description

Organism: *Rattus norvegicus* (B cell); *Mus musculus* (myeloma), rat (B cell); mouse (myeloma)

Isotype: IgM

Cell Type: hybridoma: B lymphocyte

Morphology: lymphoblast

Growth Properties: suspension

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C , preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C . Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately $125 \times g$ for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio) and dispense into a new culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product.

Handling Procedure for Flask Cultures

Recommended Handling Upon Receipt:

Suspension Cultures: The culture flask was seeded, see batch data above, and completely filled with medium to prevent loss of cells in transit. Upon receipt incubate the flask in an upright position for several hours to return the flask contents to 37°C . After the temperature has equilibrated, aseptically remove the entire contents of the flask and centrifuge at $300 \times g$ for 15 minutes. Resuspend the cell pellet in 10-12 mL of the shipping medium. From this suspension remove a sample for a cell count and viability so that the cell density of the suspension can be adjusted to $2-5 \times 10^5$ viable cells/mL. If the suspension needs to be diluted use the shipping medium. Incubate the culture in a flat position at 37°C in a 5% CO_2 in air atmosphere. Maintain the cell density of the culture as suggested under the subculture procedure described above.

Subculturing Procedure

Medium Renewal: Every 2 to 3 days



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Cultures can be maintained by addition or replacement of fresh medium. Start cultures at 2×10^5 cells/mL and maintain between 1×10^5 and 1×10^6 cells/mL.

Comments

Animals were immunized with mouse lymph node stroma.

Spleen cells were fused with Sp2/0-Ag14 myeloma cells.

The antibody specifically blocks lymphocyte adhesion to mouse peripheral lymph node endothelia and does not block lymphocyte binding to mouse Peyer's patches.

It binds to a sulfate and fucose dependent epitope associated with L-selectin binding ligands of high endothelial venules (HEV).

It inhibits L-selectin dependent lymphocyte homing to lymph nodes and blocks lymphocyte binding to HEV in vitro.

This line grows best when cultured with a feeder layer of mouse (BALB/c) peritoneal exudate cells.

References

References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S.

Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Disclaimers

This product is intended for laboratory research purposes only. It is not intended for use in humans.

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.

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