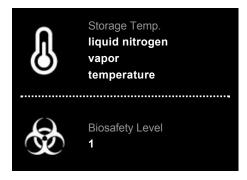


Product Sheet

WBC264-9C (ATCC[®] HB-8902[™])

Please read this FIRST



Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

The base medium for this cell line is ATCC-formulated Eagle's Minimum Essential Medium, Catalog No. 30-2003. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: WBC264-9C (ATCC $^{\otimes}$ HB-8902 $^{\text{TM}}$)

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Q Desc

Description

Organism: human (leukocyte); mouse (macrophage)

Strain:

Strain: BALB/c (macrophage)

Tissue: peripheral blood

Cell Type: Macrophage, Macrophage-like

Morphology: macrophage
Growth Properties: adherent

Cytogenetic Analysis: at passage 6, the mean number of chromosome was 64 with a range of 36 to 81



Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.



SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.



Unpacking & Storage Instructions

- 1. Check all containers for leakage or breakage.
- Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.



Handling Procedure for Frozen Cells

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at

-70°C will result in loss of viability.

SAFETY PRECAUTION: ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

- 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by
 dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under
 strict aseptic conditions.
- 3. Transfer the vial contents to a 75 cm² tissue culture flask and dilute with the recommended complete culture medium (see the specific batch information for the recommended dilution ratio). It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
- 4. Incubate the culture at 37° C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

It is not necessary to remove the cryoprotective agent. If it is desired that the cryoprotective agent be removed immediately, or that a more concentrated cell suspension be obtained, centrifuge the cell suspension at approximately 125 xg for 5 to 10 minutes. Discard the supernatant and resuspend the cells with fresh growth medium at the dilution ratio recommended in the specific batch information.



Handling Procedure for Flask Cultures

Handling Procedure for Flask Cultures (Monolayer)

The flask was seeded with cells (see specific batch information) grown and completely filled with medium at ATCC to prevent loss of cells during shipping.

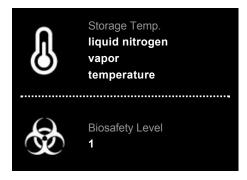
Upon receipt visually examine the culture for macroscopic evidence of any microbial contamination.



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Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination. Also check to determine if the majority of cells are still attached to the bottom of the flask; during shipping the cultures are sometimes handled roughly and many of the cells often detach and become suspended in the culture medium (but are still viable).

- If the cells are still attached, aseptically remove all but 5 to 10 mL of the shipping medium. The
 shipping medium can be saved for reuse. Incubate the cells at 37°C in a 5% CO₂ in air atmosphere until they
 are ready to be subcultured.
- 3. **If the cells are not attached,** aseptically remove the entire contents of the flask and centrifuge at 125 xg for 5 to 10 minutes. Remove shipping medium and save. Resuspend the pelleted cells in 10 mL of this medium and add to 25 cm 2 flask. Incubate at 37°C in a 5% CO $_2$ in air atmosphere until cells are ready to be subcultured.



Subculturing Procedure

Protocol: Subcultures are prepared by scraping. DO NOT USE TRYPSIN! Remove old medium, add fresh, dislodge cells and dispense into new flasks.

Subcultivation Ratio: A subcultivation ratio of 1:2 to 1:6 is recommended

Medium Renewal: Every 2 to 3 days



Cryopreservation Medium

Cryoprotectant Medium

Complete culture medium described above supplemented with 5% (v/v) DMSO. Cell culture tested DMSO is available as ATCC Catalog No. 4-X.



Comments

This is a macrophage-like cell line derived by fusion of normal human peripheral blood leukocytes with the mouse macrophage cell line, RAW 264.

The cells exhibit chemotaxis to N-formyl-methionine-leucine-phenylalanine (fMet-Leu-Phe, FMLP). WBC264-9 cells express human HPRT.



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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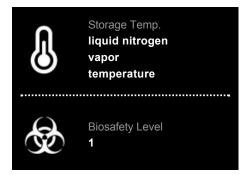
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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.

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