



Product Sheet

FHCR-1-2075/FH 4 MOUSE HYBRIDOMA

(HB-8775)

Please read this FIRST



Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

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U.S. Patent Number:
4,876,199

Technical Information

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Product Description

Designation: FHCR-1-2075/FH 4 MOUSE HYBRIDOMA
Organism: *Mus musculus* (B cell); *Mus musculus* (myeloma)
, mouse (B cell); mouse (myeloma)
Isotype: IgG3
Cell Type: hybridoma: B lymphocyte
Morphology: lymphoblast
Growth Properties: suspension

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

Part A. FROZEN CELLS

Vol./Ampule: 1.0 mL.

Recommended Handling Upon Receipt: Initiate culture as soon as possible upon receipt. Thaw by rapid agitation in 37°C water bath. See instructions on back.

Dilute ampule contents with culture medium (see batch data above). The depositor recommends the use of fresh thymocytes as a feeder layer when initiating the cells from a thawed ampule into culture.

Handling Procedure for Flask Cultures

Part B. FLASK CULTURES

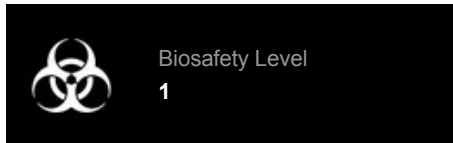
Suspension Cultures: The culture flasks have been completely filled with medium for shipment. Remove the entire contents of the flask and centrifuge at 300 x g for 15 minutes. Resuspend the cell pellet as suggested under subculture procedure described above.



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Subculturing Procedure

Medium Renewal: Every 2 to 3 days

Cultures can be maintained by addition or replacement of fresh medium. Start cultures at 5×10^4 cells/ml and maintain between 1×10^5 and 2×10^6 cells/ml. The line is best carried as an ascites in pristane primed BALB/c mice.



Comments

Animals were immunized with glycolipids adsorbed to *Salmonella minnesota*.

Spleen cells were fused with Sp2/0 myeloma cells.

The antibody is specific for di- and tri- fucosylated type 2 chain glycolipids, but does not react with monofucosylated type 2 chains.

A feeder layer of mouse peritoneal macrophages or X-irradiated STO cells (see ATCC 56-X) is necessary for successful initiation of a culture from a thawed ampule.



Propagation

Complete Growth Medium

RPMI 1640 medium, 85%; fetal bovine serum, 15% - OR - Modified Dulbecco's medium, 85%; fetal bovine serum, 15%

Culture Medium: RPMI medium 1640, 85%; fetal bovine serum, 15%.



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S.

Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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