**Product Sheet**

**U-CH17S (ATCC® CRL-3401™)**

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**Description**

- **Organism:** Homo sapiens, human
- **Tissue:** skin metastasis from a primary sacral chordoma
- **Disease:** chordoma; malignant
- **Age:** 38
- **Gender:** male
- **Morphology:** fibroblast
- **Growth Properties:** adherent

**DNA Profile:**

- Amelogenin: X,Y
- CSF1PO: 12,14
- D13S317: 11
- D16S539: 13
- DSS818: 12
- D7S820: 9,12
- TH01: 9,3
- TPOX: 11
- vWA: 14,17

**Intended Use**

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

**Complete Growth Medium**

The base medium for this cell line is Iscove’s Modified Dulbecco’s Medium (IMDM; ATCC® CRL-30-2005). RPMI-1640 Medium (ATCC® CRL-30-2001) at a 4 to 1 ratio. To 500 mL IMDM/RPMI 1640 (4:1) add the following components to make the complete medium:

- 56 mL FBS (ATCC® CRL-30-2020), final concentration of 10%
- 5.6 mL L-glutamine from stock 200 mM (ATCC® CRL-30-2214), final concentration of 1%

**Citation of Strain**

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: U-CH17S (ATCC® CRL-3401™)

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**SAFETY PRECAUTION**

ATCC highly recommends that protective gloves and clothing always be used and a full face mask be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

**Unpacking & Storage Instructions**

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

**Handling Procedure for Frozen Cells**

To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium, and spin at approximately 150-400g x g for 8 to 12 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6), pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

**Subculturing Procedure**

Volumes used in this protocol are for 75 cm² flask; proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes. Corning® T-75 flasks (catalog #430641) are recommended for subculturing this product.

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin-0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessels.
   Cultures can be established between 2.0 x 10⁴ and 6.0 x 10⁴ viable cells/cm².
6. Incubate cultures at 37°C.

Interval: Maintain cultures at a cell concentration between 2.0 x 10⁴ and 1.5 X 10⁵ cell/cm².

Subcultivation Ratio: A subcultivation ratio of 1:4 is recommended

Medium Renewal: 2 to 3 times per week

Cryopreservation Medium

Complete culture media + 5% DMSO (ATCC 4-X)

Comments

Tumor type/stage/grade: pT2b, V0, L0, Pn), N0(0/3), M1

References

References and other information relating to this product are available online at www.atcc.org.

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This product is intended for laboratory research purposes only. It is not intended for use in humans.

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Additional information on this culture is available on the ATCC web site at www.atcc.org.

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