**Description**

**Organism:** Homo sapiens, human  
**Tissue:** peritoneal effusion; derived from metastatic site: peritoneal cavity  
**Disease:** large cell lymphoma; diffuse mixed histiocytic and lymphocytic lymphoma; follicular B cell lymphoma  
**Cell Type:** B lymphocyte  
**Gender:** male  
**Morphology:** lymphoblast-like  
**Growth Properties:** suspension  

**DNA Profile:**
- DSS818: 12
- D13S317: 12,14
- D7S820: 10
- D16S539: 11,12
- vWA: 14,17
- TH01: 6,9,3
- TPOX: 11,12
- CSF1PO: 10

**Amelogenin:** X

**Cyto genetic Analysis:** This is a pseudodiploid cell line with a modal chromosome number of 47, a polyploidy rate of approximately 9% and one copy of the X chromosome. No Y chromosome was observed in any of the analyzed cells. A derivative t(14;18)(q32;q21) chromosome was present in most of the examined cells. Other derivative chromosomes were generally consistent with previous studies, these include: del(6)(p23), del(9)(p21.1?), add(11)(q25), i(17q) [or t(6?;17)(p10;q10)?] and del(18)(q21)?: [PubMed: 3881165]

**Unpacking & Storage Instructions**

**Handling Procedure for Frozen Cells**

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio) and dispense into a new culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product.

**Handling Procedure for Flask Cultures**

**SAFETY PRECAUTION**

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

**Citation of Strain**

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: SU-DHL-6 (ATCC® CRL-2959™)
SU-DHL-6 (ATCC® CRL-2959™)

Storage Temp. liquid nitrogen vapor phase

Biosafety Level 1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

The base medium for this cell line is ATCC-formulated RPMI-1640 Medium, ATCC 30-2001. To make the complete growth medium, add the following components to the base medium: fetal bovine serum (ATCC 30-2020) to a final concentration of 10%.

Citation of Strain

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Subculturing Procedure

Cultures can be maintained by the addition of fresh medium. An inoculum of 8 x 10^4 to 3 x 10^5 cells/mL is recommended. Subculture when cell concentration is between 7 x 10^5 and 1 x 10^6 cells/mL.

Subcultivation ratio: A subcultivation ratio of 1:3 to 1:6 is recommended.

Medium renewal: Every 3 to 4 days

Cryopreservation Medium

Complete growth medium with extra 20% FBS and 10% DMSO. Cell culture tested DMSO is available as ATCC Catalog No. 4-X.

Comments

All of the DHL cell lines were negative for the presence of Epstein-Barr virus (EBV) genomes. SU-DHL-6 possesses a t(14;18)(q32;q21) translocation and demonstrates an unexpected recombination within its heavy chain gene locus that may be the interchromosomal breakpoint. In SU-DHL-6, the t(14;18) translocation juxtaposes a truncated bcl-2 gene with J6 in a tail-to-head configuration. The deregulated expression of the altered bcl-2 gene may play a critical role in the disordered growth and differentiation of follicular B cell lymphoma.

References

References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the Biosafety in Microbiological and Biomedical Laboratories from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Disclaimers

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product. The MTA is also available on our Web site at [www.atcc.org](http://www.atcc.org).

Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).

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