Product Sheet

HuZP3-CHOLec3.2.8.1
(ATCC® CRL-2866™)

Please read this FIRST

Storage Temp.
liquid nitrogen vapor phase

Biosafety Level 2

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

Alpha minimum essential medium without ribonucleosides and deoxyribonucleosides, 90%; fetal bovine serum, 10%

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: HuZP3-CHOLec3.2.8.1 (ATCC® CRL-2866™)

SAFETY PRECAUTION

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

SAFETY PRECAUTION: ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Handling Procedure for Frozen Cells

1. Transiently store the vial at temperatures below -130°C, preferably in liquid nitrogen vapor, until ready for use.
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 ml complete culture medium and spin at approximately 125 x g for 5 to 10 minutes.
4. Resuspend the vial contents in the recommended complete medium (see the specific batch information for the recommended dilution ratio) and dispense into a 25 cm² or a 75 cm² culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the cell contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product.

Subculturing Procedure

Protocol: Volumes used in this protocol are for 75 sq cm flasks; proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes.
1. Remove and discard culture medium.
2. Briefly rinse the cell layer with Ca²⁺/Mg²⁺ free Dulbecco's phosphate-buffered saline (D-PBS) or 0.25% (w/v) Trypsin - 0.53 mM EDTA solution to remove all traces of serum which contains trypsin inhibitor.
3. Add 1.0 to 2.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting.

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

Description

Organism: Cricetulus griseus, hamster, Chinese
Tissue: ovary
Gender: female
Morphology: epithelial
Growth Properties: adherent
5. Transfer cell suspension to a centrifuge tube and spin at approximately 125 X g for 5 to 10 minutes. Discard supernatant.
6. Resuspend the cell pellet in fresh growth medium. Add appropriate aliquots of the cell suspension to new culture vessels. An inoculum of 3 X 10(3) to 5 X 10(3) viable cells/sq. cm is recommended.
7. Incubate cultures at 37°C. We recommend that you maintain cultures at a cell concentration between 8 X 10(4) and 1.5 X 10(5) cells/sq. cm.

Subcultivation Ratio: A subcultivation ratio of 1:4 to 1:8 is recommended

Medium Renewal: Every 2 to 3 days

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### Cryopreservation Medium

**Cryoprotectant Medium**

Complete growth medium described above supplemented with 5% (v/v) DMSO.

Cell culture tested DMSO is available as ATCC® Catalog No. 4-X.

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### Comments

HuZP3-CHOlec3.2.8.1 cells express recombinant human ZP3. The zona pellucida is an extracellular matrix that mediates taxon-specific fertilization and in the mouse is composed of three glycoproteins (ZP1, ZP2, ZP3). Full length human ZP3 cDNA was cloned into expression vector pEF1/V5-His and transfected into glycosylation-deficient CHO cells. It was compared to mouse ZP3 and found to be quite similar [PubMed: 15379548].

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### References

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).

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### Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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### Disclaimers

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Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).

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