



Product Sheet

WT 9-12 (ATCC® CRL-2833™)

Please read this **FIRST**



Storage Temp.
liquid nitrogen
vapor phase



Biosafety Level
2

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

The base medium for this cell line is ATCC-formulated Dulbecco's Modified Eagle's Medium, Catalog No. 30-2002. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: WT 9-12 (ATCC® CRL-2833™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor

Description

Organism: *Homo sapiens*, human
Tissue: kidney; cyst from a distal and proximal cortical tubule
Disease: autosomal dominant polycystic kidney disease (ADPKD)
Cell Type: epithelial
Age: unknown
Gender: female
Morphology: polygonal
Growth Properties: adherent

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

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NOTE: Flasks must be coated with **Bovine Collagen type I** solution (supplied stock concentration is 3.0 mg/mL; PureCol® Advanced Biomatrix Catalog No. 5005-B) for 1 hour at 37°C

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 ml complete culture medium and spin at approximately 125 xg for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio) and dispense into a 25 cm² or a 75 cm² culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product.

Subculturing Procedure

Protocol:

Flasks must be coated with Bovine Collagen type I solution (supplied stock concentration is 3.0 mg/mL; PureCol® Advanced Biomatrix Catalog No. 5005-B) for 1 hour at 37°C.


1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin - 0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.




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3. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37C to facilitate dispersal.
4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new Bovine Collagen Type I coated flasks.
An inoculum of 3 to 5 X 10³ viable cells/sq. cm is recommended.
6. Incubate cultures at 37C.

Interval: Maintain cultures at a cell concentration between 5 X 10³ and 8 X 10⁴ cells/sq. cm

Subcultivation Ratio: A subcultivation ratio of 1:4 to 1:8 is recommended

Medium Renewal: Two to three times weekly



Cryopreservation Medium

Cryoprotectant Medium

Complete growth medium described above supplemented with 5% (v/v) DMSO.
Cell culture tested DMSO is available as ATCC Catalog No. 4-X.



Comments

The WT 9-12 cell line was established in July, 1999 from a single cyst from a cortical tubule taken from a patient with autosomal dominant polycystic kidney disease (ADPKD). The cells have both distal and proximal characteristics. They can be subcloned to obtain clones with proximal or distal markers. The cells were transformed with a wild type recombinant adeno-SV40 virus. These viruses contain an origin-deficient (ori-) WT SV40 DNA cloned into the adenovirus vector in place of regions 1a and 1b [PubMed: 12734101]. WT 9-12 is one of a series of immortalized epithelial cell lines from over 30 individual renal cysts obtained from 11 patients with ADPKD. These new series of cyst-derived cell lines represent useful in vitro models for studying the cellular and molecular biology of ADPKD.

Antigen: SV40 large T antigen, expressed [PubMed: 12734101]



References

References and other information relating to this product are available online at www.atcc.org.



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Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

ATCC Warranty

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Disclaimers

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org




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