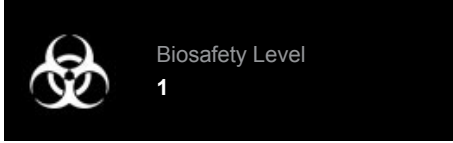




Product Sheet

R2-9A5 [R2-9A5.2A2.1A10] (ATCC® CRL-2357™)

Please read this FIRST



Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

Iscove's Modified Dulbecco's Medium with 4 mM L-glutamine adjusted to contain 1.5 g/L sodium bicarbonate supplemented with 0.05 mM 2-mercaptoethanol and 100 U/ml recombinant human IL-6, 90%; fetal bovine serum, 10%.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: R2-9A5 [R2-9A5.2A2.1A10] (ATCC® CRL-2357™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor

Description

Organism: *Rattus norvegicus* (B cell); *Mus musculus* (myeloma), rat (B cell); mouse (myeloma)
Isotype: IgG2b
Cell Type: hybridoma: B lymphocyte
Growth Properties: suspension

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

Recommended Handling Upon Receipt: Initiate culture as soon as possible upon receipt. Thaw by rapid agitation in 37°C water bath. See instructions on back. Dilute ampule contents with culture medium (see batch data above). Add fresh medium as density increases.

Handling Procedure for Flask Cultures

Suspension Cultures: The culture flask was seeded, see batch data above, and completely filled with medium to prevent loss of cells in transit. Upon receipt incubate the flask in an upright position for several hours to return the flask contents to 37°C. After the temperature has equilibrated, aseptically remove the entire contents of the flask and centrifuge at 300 x g for 15 minutes. Resuspend the cell pellet in 10-12 ml of the shipping medium. From this suspension remove a sample for a cell count and viability so that the cell density of the suspension can be adjusted to 2-5 x 10⁵ viable cells/ml. If the suspension needs to be diluted use the shipping medium. Incubate the culture in a flat position at 37°C in a 5% CO₂ in air atmosphere. Maintain the cell density of the culture as suggested under the subculture procedure described above.

Subculturing Procedure

Medium Renewal: Every 2 to 3 days

Cultures can be maintained by addition or replacement of fresh medium. Alternately, cultures can be established by centrifugation with subsequent resuspension in fresh culture medium.

Establish new cultures at 2 X 10⁵ viable cells/ml and maintain at between 1 X 10⁵ and 1 X 10⁶ viable cells/ml.

Cryopreservation Medium

Culture medium, 95%; DMSO, 5%.

Comments

The hybridoma cell line R2-9A5 [R2-9A5.2A2.1A10] was established by David Presky and Victoria Wilkinson in 1993 and subcloned twice.

The antibody is a useful tool for detection of mouse IL-12p75 by ELISA and Western Blot analysis.

R2-9A5 was formed by the fusion of SP2/0 mouse myeloma cells with splenocytes from an adult female Lewis rat inoculated with recombinant mouse IL-12 p75.

References



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References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Disclaimers

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.

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