



Product Sheet

RPE-J (ATCC® CRL-2240™)

Please read this FIRST



Storage Temp.
liquid nitrogen
vapor phase



Biosafety Level
2

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

Dulbecco's modified Eagle's medium with 4 mM L-glutamine adjusted to contain 1.5g/L sodium bicarbonate and 4.5 g/L glucose and supplemented with 0.1 mM non-essential amino acids, 96%; fetal bovine serum, 4%

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: RPE-J (ATCC® CRL-2240™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor

Description

Organism: *Rattus norvegicus*, rat
Strain: Long-Evans
Tissue: eye; retinal pigmented epithelium; retina
Disease: normal
Cell Type: epithelial SV40 transformed
Age: 7 days
Morphology: epithelial
Growth Properties: adherent

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 33°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio) and dispense into a 25 cm² or a 75 cm² culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 33°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Handling Procedure for Flask Cultures

The flask was seeded with cells (see specific batch information), grown, and completely filled with medium at ATCC to prevent loss of cells during shipping.

1. Upon receipt, visually examine the culture for macroscopic evidence of any microbial contamination. Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination. Also, check to determine if the majority of cells are still attached to the bottom of the flask; during shipping the cultures are sometimes handled roughly and many of the cells often detach and become suspended in the culture medium (but are still viable).
2. **If the cells are still attached**, aseptically remove all but 5 to 10 mL of the shipping medium. The shipping medium can be saved for reuse. Incubate the cells at 33°C in a 5% CO₂ in air atmosphere until they are ready to be subcultured.
3. **If the cells are not attached**, aseptically remove the entire contents of the flask and centrifuge at 125 x g for 5 to 10 minutes. Remove shipping medium and save. Resuspend the pelleted cells in 10



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mL of this medium and add to 25 cm² flask. Incubate at 33°C in a 5% CO₂ in air atmosphere until cells are ready to be subcultured.



Subculturing Procedure

Volumes used in this protocol are for 75 cm² flask; proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes.

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin-0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting.
5. Transfer cell suspension to a centrifuge tube and spin at approximately 125 x g for 5 to 10 minutes. Discard supernatant.
6. Resuspend the cell pellet in fresh growth medium. Add appropriate aliquots of cell suspension to new culture vessels.
7. Place culture vessels in incubators at 33°C.

Subculture Ratio: 1:3

Medium Renewal: Every 2 to 3 days.

Note: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 13 in **Culture of Animal Cells, a manual of Basic Technique** by R. Ian Freshney, 5th edition, published by Wiley-Liss, N.Y., 2005.



Cryopreservation Medium

Complete growth medium (see above) with an additional 16% fetal bovine serum and 5% (v/v) DMSO. Cell culture tested DMSO is available as ATCC® Catalog No. 4-X.



Comments

The cells express a transformed phenotype at the permissive temperature (33°C), and a non-transformed phenotype at the non-permissive temperature (40°C). They must be cultured at the permissive temperature and do not grow at 37°C.

When RPE-J cells are grown on nitrocellulose filters coated with a thin layer of Matrigel in the presence of 10(-8) M retinoic acid for 6 days at 33°C and then switched to the non-permissive temperature of 40°C for 33 to 36 hours, they acquire a differentiated polarized RPE phenotype. Under these conditions, RPE-J cells exhibit circumferential staining for the tight-junction protein ZO-1 and acquire a transepithelial resistance of 350 ohms/cm². [Ref](#)

RPE-J is the only established RPE cell line that maintains epithelial cell surface polarity. The cells retain many properties of RPE including expression of the rat RPE marker RET-PE2 and the ability to phagocytose latex beads. [Ref](#)

A culture submitted to the ATCC in July 1995 was found to be contaminated with Mycoplasma hyorhinis and was cured by a 21-day treatment with BM Cycline. The cells were assayed for mycoplasma by the Hoechst stain and the standard culture test over a six-week period following treatment and all tests were negative. RPE-J is a retinal pigment epithelial (RPE) cell line derived from primary cultures of RPE cells taken from 7-day-old Long-Evans rats.



References

References and other information relating to this product are available online at www.atcc.org.



Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.



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Disclaimers

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