Product Sheet
DT40 (ATCC® CRL-2111™)

Please read this FIRST

Storage Temp.
liquid nitrogen
vapor phase

Biosafety Level
1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

Dulbecco's modified Eagle's medium with 4 mM L-glutamine modified to contain 4.5 g/L glucose, 1.5 g/L sodium bicarbonate and 0.05 mM 2-mercaptoethanol, 75%; tryptose phosphate broth, 10%; fetal bovine serum, 10%; chicken serum, 5%

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: DT40 (ATCC® CRL-2111™)

Description

Organism: Gallus gallus, chicken
Tissue: bursa
Disease: lymphoma
Cell Type: lymphoblast
Age: 1 day
Morphology: lymphoblast
Growth Properties: suspension

Batch-Specific Information

Refer to the Certificate of Analysis for batch-specific test results.

SAFETY PRECAUTION

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

   1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
   2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
   3. It is recommended that the cryoprotective agent be removed immediately. Centrifuge the cell suspension at approximately 125 x g for 5 to 10 minutes. Discard the supernatant and resuspend the cell pellet in an appropriate amount of fresh growth medium.
   4. Transfer the cell pellet to an appropriate size vessel. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
   5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Handling Procedure for Flask Cultures

The flask was seeded with cells (see specific batch information), grown, and completely filled with medium at ATCC to prevent loss of cells during shipping.

   1. Upon receipt visually examine the culture for macroscopic evidence of any microbial contamination. Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination
   2. Incubate the flask in an upright position for several hours at 37°C. After the temperature has equilibrated, aseptically remove the entire contents of the flask and centrifuge at 125 x g for 5 to 10 minutes. Remove shipping medium and save for reuse. Resuspend the cell pellet in 10 mL of this medium.
   3. From this cell suspension remove a sample for a cell count and viability. Adjust the cell density of the suspension to 3 x 10⁶ viable cells/mL in the shipping medium.
   4. Incubate the culture, horizontally, at 37°C in a 5% CO₂ in air atmosphere. Maintain the cell density of the culture as suggested under the subculture procedure.

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org
800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

Or contact your local distributor
Subculturing Procedure

Cultures can be maintained by addition or replacement of fresh medium. Start cultures at $3 \times 10^5$ cells/mL and maintain between $2 \times 10^5$ and $2 \times 10^6$ cells/mL.

Medium Renewal: Every 2 to 3 days

Cryopreservation Medium

Complete growth medium described above supplemented with 5% (v/v) DMSO.

Cell culture tested DMSO is available as ATCC Catalog No. 4-X.

Comments

The cell line contains proviral DNA sequences integrated upstream from the c-myc proto-oncogene and expresses increased levels of c-myc RNA. It lacks a normal c-myc gene, but contains two copies of an ALV deregulated myc gene. The cells retain the ability to rearrange the immunoglobulin light chain gene (IgL). At the IgL locus, DT40 contains one rearranged and one germline allele.

The cell line exhibits a lymphoblastoid phenotype. Infectivity assays revealed that DT40 releases low levels of infectious RAV-1. Both the c-rel gene and the v-rel oncogenes induce major histocompatibility (MHC) class II antigen expression on the DT40 cell line. The expression of MHC class II is induced more rapidly by v-rel than c-rel and several weeks after infection, rel induced class II antigen as much as 50 fold more efficiently than c-rel.

References

References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the Biosafety in Microbiological and Biomedical Laboratories from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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