**Unpacking & Storage Instructions**

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

**Handling Procedure for Frozen Cells**

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 24°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a 75 cm² tissue culture flask and dilute with the recommended complete culture medium (see the specific batch information for the recommended dilution ratio).
4. Incubate the culture at 24°C in a suitable incubator.

If it is desired that the cryoprotective agent be removed immediately, or that a more concentrated cell suspension be obtained, centrifuge the cell suspension at approximately 125 x g for 5 to 10 minutes. Discard the supernatant and resuspend the cells with fresh growth medium at the dilution ratio recommended in the specific batch information.

**Handling Procedure for Flask Cultures**

The flask was seeded with cells (see specific batch information), grown, and completely filled with medium at ATCC to prevent loss of cells during shipping.

1. Upon receipt visually examine the culture for macroscopic evidence of any microbial contamination. Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination.
2. Aseptically remove the entire contents of the flask and centrifuge at 125 x g for 5 to 10 minutes. Remove shipping medium and save for reuse. Resuspend the cell pellet in 10 mL of this medium.
3. From this cell suspension, remove a sample for a cell count and viability. Adjust the cell density of the suspension to 2-5 x 10⁵ viable cells/mL in the shipping medium.
4. Incubate the culture, horizontally, at 24°C. Maintain the cell density of the culture as suggested under the subculture procedure.

**Subculturing Procedure**

Gently resuspend the cells by pipetting medium across the monolayer or (for larger flasks) by slapping the flask against the base of the palm. Dilute the cell suspension to the desired concentration with fresh medium. Maintain cultures at a cell concentration between 5 x 10⁴ and 4 x 10⁵ cells/cm².

**Subcultivation Ratio:** A subcultivation ratio of 1:10 or greater is recommended.
Medium Renewal: At the time of subcultivation

Note: The cells will grow as a loose monolayer (or in suspension) at any temperature between 16°C and 28°C.

Cryopreservation Medium

Complete culture medium described above supplemented with 5% (v/v) DMSO. Cell culture tested DMSO is available as ATCC Catalog No. 4-X.

Comments

The cells have been shown to support the growth of the insect stages of malaria parasites. ref

References

References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the Biosafety in Microbiological and Biomedical Laboratories from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Schneider's Drosophila Line 2 [D. Mel. (2), SL2] (ATCC® CRL-1963™)

Please read this FIRST

Storage Temp.

liquid nitrogen vapor phase

Biosafety Level

1

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium

Schneider's Drosophila Medium, 90%; heat-inactivated fetal bovine serum, 10%. NOTE: the fetal bovine serum should be selected for ability to support the growth of insect cell lines. This medium is formulated for use without CO2.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: Schneider's Drosophila Line 2 [D. Mel. (2), SL2] (ATCC® CRL-1963™)