Organism: *Homo sapiens*, human

Tissue: skin

Disease: malignant melanoma

Age: 54 years

Gender: female

Morphology: epithelial

Growth Properties: adherent

DNA Profile:
- Amelogenin: X
- CSF1PO: 11,12
- D13S317: 11,14
- D16S539: 9
- D5S818: 12
- D7S820: 9
- TH01: 8
- TPOX: 8,10
- vWA: 16,17

Refer to the Certificate of Analysis for batch-specific test results.

ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

Unpacking & Storage Instructions

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells

To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium, and spin at approximately 150 x g for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6). pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Subculturing Procedure

Volumes used in this protocol are for 75 cm² flask; proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes. Corning® T-75 flasks (catalog #430641) are recommended for subculturing this product.

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin-0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach.
4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cultures at 37°C.

**Interval:** Maintain cultures at a cell concentration between $8 \times 10^3$ and $4 \times 10^4$ cells/cm².

**Subcultivation Ratio:** A subcultivation ratio of 1:2 to 1:6 is recommended

**Medium Renewal:** 2 to 3 times per week

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**Complete Growth Medium**

The base medium for this cell line is ATCC-formulated Dulbecco's Modified Eagle's Medium (DMEM; ATCC 30-2002). To make the complete medium add the following component to the base medium: Fetal bovine serum (FBS; ATCC 30-2020) to a final concentration of 10%.

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**Citation of Strain**

This is a malignant BRAFV600E/KRASG13D mutant isogenic line derived from the parental A375 (ATCC CRL-1619) cell line. The c.38G>A knock-in mutation encoding KRAS p.G13D protein expression was generated at ATCC by utilizing the CRISPR/Cas9 gene editing technology. This is a heterozygous mutation expressing the KRAS wild-type and the c.38G>A mutant alleles.

BRAF is a proto-oncogene encoding B-RAF, a serine/threonine kinase of the RAF family that acts downstream of RAS and upstream of MEK in the MAPK/ERK signaling pathway. Mutations in BRAF lead to excessive cellular proliferation, differentiation, and survival. BRAF V600E mutations are present in 50% of melanomas and although there are current BRAF inhibitors used as successful therapeutics, patients often become resistant to drugs several months following treatment. One mechanism of resistance to these inhibitors is caused by upstream secondary RAS acquired mutations. The KRASG13D mutant isogenic line, ATCC CRL-1619IG-1 has been validated at the genomic, transcript, and protein bio-functional levels and exhibits significant resistance to the BRAF inhibitors Dabrafenib and Vemurafenib when compared to its parental cell line. CRL-1619IG-1 can be a useful model to study the RAS–RAF–MEK–ERK–MAP kinase signaling pathway and to screen potential BRAF inhibitors and anti-cancer compounds for drug discovery and development.

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**References**

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).

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**Biosafety Level: 2**

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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