Organism: *Homo sapiens*, human
Tissue: peripheral blood
Disease: acute promyelocytic leukemia
Cell Type: promyeloblast
Age: 36 years
Gender: female
Morphology: myeloblastic
Growth Properties: suspension
Isoenzymes:
- AK-1, 1
- ES-D, 1
- G6PD, B
- GLO-I, 1
- Me-2, 1
- PGM1, 1
- PGM3, 1
DNA Profile:
- D5S818: 12
- D13S317: 8,11
- D7S820: 11,12
- D16S539: 11
- vWA: 16
- TH01: 7,8
- Amelogenin: X
- TPOX: 8,11
- CSF1PO: 13,14

Cytogenetic Analysis:
The stemline chromosome number is pseudodiploid with the 2S component occurring at 6.2%. Five markers (M2 through M6) were common to most S metaphases. DM's, which varied in numbers per cell, occurred in all metaphases karyotyped. HSR chromosomes were not detected.

SAFETY PRECAUTION

Unpacking & Storage Instructions
1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Handling Procedure for Frozen Cells
To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.
1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium. and spin at approximately 125 x g for 5 to 7 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture
Please read this FIRST

Storage Temp.
liquid nitrogen vapor phase
Biosafety Level 1

Intended Use
This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Complete Growth Medium
The base medium for this cell line is ATCC-formulated Iscove's Modified Dulbecco's Medium, Catalog No. 30-2005. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 20%.

Citation of Strain
If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: HL-60 (ATCC® CCL-240™)

Handling Procedure for Flask Cultures
The flask was seeded with cells (see specific batch information), grown, and completely filled with medium at ATCC to prevent loss of cells during shipping.

1. Upon receipt visually examine the culture for macroscopic evidence of any microbial contamination. Using an inverted microscope (preferably equipped with phase-contrast optics), carefully check for any evidence of microbial contamination
2. Incubate the flask in an upright position for several hours at 37°C. After the temperature has equilibrated, aseptically remove the entire contents of the flask and centrifuge at 125 x g for 5 to 10 minutes. Remove shipping medium and save for reuse. Resuspend the cell pellet in 10 mL of this medium.
3. From this cell suspension remove a sample for a cell count and viability. Adjust the cell density of the suspension to 2 to 3 x 10^6 viable cells/mL in the shipping medium.
4. Incubate the culture, horizontally, at 37°C in a 5% CO₂ in air atmosphere. Maintain the cell density of the culture as suggested under the subculture procedure.

Subculturing Procedure
Cultures can be maintained by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 1 X 10^6 viable cells/mL. Do not allow cell concentration to exceed 1 X 10^6 cells/mL. Corning® T-75 flasks (catalog #431464) are recommended for subculturing this product.

Interval: Maintain cell density between 1 x 10^5 and 1 x 10^6 viable cells/mL.
Medium Renewal: Every 2 to 3 days

Cryopreservation Medium
Iscove’s Modified Dulbecco’s Medium, 30-2005, supplemented with 7% FBS and 5% (v/v) DMSO. Lots produced prior to May 2019 may have used a different cryopreservation medium (complete growth medium supplemented with 5% (v/v) DMSO), contact Technical Support for further details.

Comments
HL-60 cells spontaneously differentiate and differentiation can be stimulated by butyrate, hypoxanthine, phorbol myristic acid (PMA, TPA), dimethylsulfoxide (DMSO, 1% to 1.5%), actinomycin D, and retinoic acid. The cells exhibit phagocytic activity and responsiveness to chemotactic stimuli. The line is positive for myc oncogene expression.

References
References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 1

ATCC Warranty
ATCC® products are warranted for 30 days from the date of shipment, and this warranty is valid only if the product is stored and handled according to the information included on this product information sheet. If the ATCC® product is a living cell or microorganism, ATCC lists the media formulation that has been found to be effective for this product. While other, unspecified media may also produce satisfactory results, a change in media or the absence of an additive from the ATCC recommended media may affect recovery, growth and/or function of this product. If an alternative medium formulation is used, the ATCC warranty for viability is no longer valid.
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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.