



Product Sheet

Helicobacter pylori (ATCC® BAA-945™)

Please read this FIRST



Storage Temp.
Frozen: -80°C or colder
Freeze-Dried: 2°C to 8°C
Live Culture: See Propagation Section



Biosafety Level
2

Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Helicobacter pylori* (ATCC® BAA-945™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
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Or contact your local distributor

Description

Designation: Baylor Challenge Strain 100 (BCS 100)

Propagation

Medium

ATCC® Medium 18: Trypticase Soy Agar/Broth

ATCC® Medium 260: Trypticase soy agar/broth with defibrinated sheep blood

Growth Conditions

Temperature: 37°C

Atmosphere: Microaerophilic

Propagation Procedure

1. This organism is shipped frozen in dry ice. Just prior to use, thaw vial in water at approximately 37°C. When thawed, a drop of the suspension may be used to do an immediate wet mount to observe the unique morphology of this organism and verify its viability by checking for motility.
2. Aseptically transfer the thawed suspension into a fresh #18 broth (3-5 mL). Mix well. This suspension can now be used to inoculate agar slant(s), plate(s), or the preferred biphasic culture. Two #260 plates should be inoculated, one for microaerophilic growth and the second for aerobic growth. No growth should occur on the plate incubated aerobically.
3. To obtain a biphasic culture, add 0.6 mL of the suspension to a # 260 slant. The resulting pool at the bottom of the slant is where the best, most rapid growth will occur.
4. Incubate at 37°C under microaerophilic conditions using an anaerobe jar with an active catalyst and a microaerophilic gas generator pack, or other acceptable method, to obtain microaerophilic conditions. An oxygen concentration of 6% is ideal. Incubate tubes with cap loose.
5. Within 3 days, good growth should be obtained in the broth pool at the bottom of the slant. Additional incubation may be required for colonies to appear on agar plate. Further subcultures can be made using the broth pool as the inoculum source. Subcultures to biphasic cultures will require only 24 to 48 hours of incubation for good growth.

Notes

This is a slow growing organism that requires moist conditions for best growth. Growth at the broth/agar interface of the biphasic slant should occur within three days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy. The organism is a medium size, regular to slightly curved, motile bacillus. Motility is usually observed only in young cultures. The presence of spheroid cells indicates that viability is being lost either due to age or too much exposure to oxygen. Once good growth is present, these organisms tend to lose viability, especially if exposed to air for lengthy periods. Viability also decreases with repeated subculturing. The cells do not Gram stain well using traditional procedures. To obtain the best results, use a basic fuchsin counterstain in place of the safranin. Once good growth is obtained, transfer or freeze the culture. Adding an equal amount of 20% sterile glycerol to pooled broth from several biphasic slants, followed by freezing in liquid nitrogen or "ultra-low temperature" freezer is recommended.

Additional information on this culture is available on the ATCC® web site at www.atcc.org.

References

References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

ATCC Warranty


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media or the absence of an additive from the ATCC recommended media may affect recovery, growth and/or function of this product. If an alternative medium formulation is used, the ATCC warranty for viability is no longer valid.

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

Additional information on this culture is available on the ATCC web site at www.atcc.org.

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