

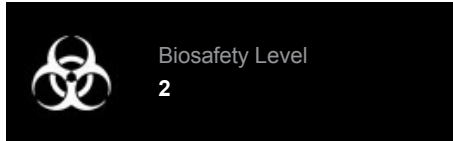


Product Sheet

## *Campylobacter jejuni*

### *subsp. jejuni* (ATCC® BAA-529™)

Please read this **FIRST**



#### Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

#### Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Campylobacter jejuni subsp. jejuni* (ATCC® BAA-529™)

#### Description

**Designation:** INP59

**Antigenic Properties:** Serotype HS:41

#### Propagation

##### Medium

ATCC® Medium 260: Trypticase soy agar/broth with defibrinated sheep blood

ATCC® Medium 18: Trypticase Soy Agar/Broth

##### Growth Conditions

**Temperature:** 37°C

**Atmosphere:** Microaerophilic (10% CO<sub>2</sub>, 3-5% O<sub>2</sub>)

##### Propagation Procedure

1. Open vial according to enclosed instructions.
2. Using a single tube of #1115 or #177 broth (5 to 6 ml), withdraw approximately 0.5 to 1.0 ml with a Pasteur or 1.0 ml pipette. Rehydrate the pellet.
3. Aseptically transfer this aliquot back into the broth tube. Mix well.
4. Use several drops of the suspension to inoculate a #260 slant, and/or plate.
5. Or, to obtain a biphasic culture, add 0.5 ml of the suspension to a #260 agar slant (see notes).
6. Incubate tubes and plate at 37°C, under microaerophilic conditions, for 24 to 48 hours. Use an anaerobe jar with an active catalyst and a microaerophilic gas generator pack, or other acceptable method. Incubate slant with cap loose.

#### Notes

Fluid Thioglycollate tube may be incubated aerobically.

Colonies are small, entire, gray, and glistening.

This is a slow growing organism that requires moist conditions for best growth. A biphasic culture will give the most rapid growth. Growth at the broth/agar interface of the biphasic slant should occur within two to three days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy. The organism is a curved to spiral shaped, motile rod. Motility is usually observed only in young cultures.

Growth on agar takes longer than with the biphasic culture. Once good growth is present, these organisms tend to lose viability, especially if exposed to air for lengthy periods.

The cells do not Gram stain well using traditional procedures. To obtain the best results, use a basic fuchsin counterstain in place of the safranin.

Storage at liquid nitrogen temperatures, with 10% sterile glycerol as the cryoprotectant, is recommended for long-term preservation.

Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).

#### References

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).

#### Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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American Type Culture Collection  
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Manassas, VA 20108 USA  
[www.atcc.org](http://www.atcc.org)

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Fax: 703.365.2750  
Email: [Tech@atcc.org](mailto:Tech@atcc.org)

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Manassas, VA 20108 USA  
[www.atcc.org](http://www.atcc.org)

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Fax: 703.365.2750  
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