**Campylobacter helveticus**  
(ATCC® 51209™)

### Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

### Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Campylobacter helveticus* (ATCC® 51209™)

### Propagation

#### Medium

- ATCC® Medium 1115: Brucella albimi broth
- ATCC® Medium 177: Fluid thioglycollate medium
- ATCC® Medium 260: Tryptase soy agar/broth with defibrinated sheep blood

#### Growth Conditions

- **Temperature:** 37°C
- **Atmosphere:** Microaerophilic gas mixture, 6% O₂

#### Propagation Procedure

1. Open vial according to enclosed instructions or visit www.atcc.org for instructions.
2. Rehydrate the entire pellet with approximately 0.5 mL of #1115 or #177 broth.
3. Aseptically transfer the entire contents to a 5-6 mL tube of #1115 or #177 broth. Additional test tubes can be inoculated by transferring 0.5 mL of the primary broth tube to these secondary broth tubes.
4. Use several drops of the primary broth tube to inoculate a #260 agar slant.
5. Incubate at 37°C under microaerophilic conditions for 48 hours. Use an anaerobe jar with an active catalyst and a microaerophilic gas generator pack or other acceptable method. An atmosphere containing a 6% oxygen concentration is ideal. All tubes and slants should be incubated with caps loosened.

#### Notes

This is a slow growing organism that requires moist conditions for best growth. A biphasic culture will give the most rapid growth. Growth at the broth/agar interface of the biphasic slant should occur within two to three days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy.

Establish growth in biphasic culture slant for 48 to 96 hours before transferring to agar plate. Growth on agar takes longer (between 5 to 7 days) after transferring from biphasic culture. Once good growth is present, these organisms tend to lose viability, especially if exposed to air for lengthy periods. Fluid Thioglycollate tubes may be incubated aerobically. The cells do not Gram stain well using traditional procedures. To obtain the best results, use a basic fuchsin counterstain in place of the safranin.

Storage at vapor phase of liquid nitrogen temperatures, with 10% sterile glycerol as the cryoprotectant, is recommended for long-term preservation.

Additional information on this culture is available on the ATCC® web site at www.atcc.org.

### References

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).

### Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org.

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