**Leishmania braziliensis**

**Strain Designation:** MHOM/BR/75/M2903

**Deposited Name:** *Leishmania braziliensis braziliensis* Vianna

**Depositor:** WRAIR

**Isolation:** Human, Serra das Carajas, Para, Brazil, 1975

## Growth Conditions

**Temperature:** 25°C

**Culture System:** Axenic

**Medium**

- ATCC® Medium 431: Trypanosome medium
- ATCC® Medium 1011: Diphasic blood agar medium
- ATCC® Medium 1012: Diphasic blood agar medium
- ATCC® Medium 2736: M199, Modified Medium
- ATCC® Medium 807: Brain heart infusion blood agar

### Instructions for Complete Medium

**Media:** ATCC medium 431 Trypanosome medium

**Alternate media:** ATCC medium 1011 Diphasic blood agar medium (30% rabbit blood), ATCC medium 1012 Diphasic blood agar medium (10% rabbit blood), ATCC Medium 2736 Modified M199, ATCC medium 807 Brain heart infusion blood agar (10% rabbit blood, BHI overlay) (some strains may not grow equally well in alternative media)

**Medium Note:** As an alternative to blood-agar media, pure broth ATCC Medium 2736 may be used in screw-capped tubes, incubated with caps tightened in an upright position to reduce the liquid surface area available for gas interchange.

## Storage and Culture Initiation

1. To thaw a frozen ampule, place in a 35°C water bath, until thawed (2-3 min). Immerse the ampule just sufficient to cover the frozen material. Do not agitate the ampule.
2. Immediately after thawing, aseptically transfer contents to a screw-capped borosilicate test tube containing ATCC Medium 431, or alternatively, inoculate 10 mL complete ATCC medium 2736 in a screw-capped test tube. Incubate the tube vertically at 25°C with the cap screwed on tightly.

## Culture Maintenance

1. When the culture has reached or is near peak density, invert tube 10 times and aseptically transfer a drop from a Pasteur pipette (0.05 mL) to another test tube of fresh medium.
2. Incubate the culture vertically at 25°C with the cap screwed on tightly.
3. Transfer the culture every 3-4 days as described in step 1. The transfer interval will depend on the quantity of the inoculum and the quality of the medium. This should be empirically determined by examining the culture on a daily basis until the growth cycle has stabilized.

### Note:

Some fastidious strains may not proliferate well in older growth medium. If growth is poor, wash cells in an osmotically-balanced saline solution, centrifuge to concentrate as indicated below, and use freshly-made growth media.

## Harvest and Preservation

1. Harvest cells from a culture which is at or near peak density by centrifugation at 1,300 g for 5 min.
2. Adjust concentration of cells to 2 x 10^7/mL in fresh medium.
3. While cells are centrifuging prepare a 10% (v/v) solution of sterile DMSO in fresh medium (broth). The DMSO solution when first prepared will warm up due to chemical heat. The solution should be allowed to return to room temperature prior to use.
4. Mix the cell preparation and the DMSO solution in equal portions. The final concentration will be 10^7 cells/mL and 5% (v/v) DMSO. The time from the mixing of the cell preparation and DMSO stock solution before the freezing process is begun should be no more than 15 min.
5. Dispense in 0.5 mL aliquots into 1.0-2.0 mL sterile plastic screw-capped cryules (special plastic vials for cryopreservation).
6. Place the ampules in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately -1°C/min.)
7. Store in either the vapor or liquid phase of a nitrogen refrigerator.
8. To thaw a frozen ampule, place it in a 35°C water bath such that the lip of the ampule remains above the water line. Thawing time is approximately 2 to 3 minutes. Do not agitate the ampule. Do not leave ampule in water bath after thawed.
9. Immediately after thawing, do not leave in the water bath, aseptically transfer the contents of the ampule into a fresh tube of ATCC medium 431, or alternatively, inoculate 10 mL complete ATCC medium 2736 in a screw-capped test tube.
10. Incubate vertically at 25°C with the cap screwed on tightly.
11. Maintain as described above.

References
References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the Biosafety in Microbiological and Biomedical Laboratories from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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