



Product Sheet

# *Campylobacter sputorum* *biovar sputorum* (ATCC® 49916™)

Please read this **FIRST**



## Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

## Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Campylobacter sputorum biovar sputorum* (ATCC® 49916™)

American Type Culture Collection  
PO Box 1549  
Manassas, VA 20108 USA  
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Fax: 703.365.2750  
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## Description

**Designation:** LRA 116.06.89

**Deposited Name:** *Campylobacter sputorum* subsp. *bubulus* (Florent) Veron and Chatelain

## Propagation

### Medium

ATCC® Medium 18: Trypticase Soy Agar/Broth

ATCC® Medium 18: Trypticase Soy Agar/Broth

ATCC® Medium 260: Trypticase soy agar/broth with defibrinated sheep blood

### Growth Conditions

**Temperature:** 37.0°C

**Atmosphere:** Microaerophilic

### Propagation Procedure

1. Open vial according to enclosed instructions. Rehydrate contents of vial with 0.5 ml of #18 broth.
2. To obtain a biphasic culture, add 0.4 ml of the suspension to a #260 slant. Add remaining 0.1 ml of the suspension to a #260 plate and streak for isolation.
3. Incubate at 37°C under microaerophilic conditions using an anaerobe jar with an active catalyst and a microaerophilic gas generator pack, or other acceptable method, to obtain microaerophilic conditions. Incubate slant with cap loose.
4. Within 24 hours of incubation, good growth should be obtained in the broth pool at the bottom of the slant and on agar surfaces. Further subcultures can be made using broth pool as the inoculum source.

## Notes

This organism requires moist conditions for best growth. Growth at the broth/agar interface of the biphasic slant should occur within 1-2 days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy. The organism is a medium size, regular to slightly curved, motile bacillus. Motility is usually observed only in young cultures. The presence of spheroid cells indicates that viability is being lost either due to age or too much exposure to oxygen.

Colonies on #260 are small, circular, entire, and gray. Once good growth is present, these organisms tend to lose viability, especially if exposed to air for lengthy periods. Viability also decreases with repeated subculturing. The cells do not Gram stain well using traditional procedures. To obtain the best results, use a basic fuchsin counterstain in place of the safranin.

Once good growth is obtained, transfer or freeze the culture. Adding an equal amount of 20% sterile glycerol to pooled broth from several biphasic slants, followed by freezing in liquid nitrogen or "ultra-low temperature" freezer is recommended.

Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).

## References

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).

## Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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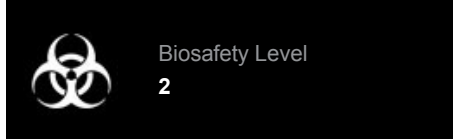


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**Disclaimers**

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