



Product Sheet

# *Chlorobaculum tepidum* (ATCC® 49652™)

Please read this **FIRST**



Storage Temp.  
**Frozen: -80°C or colder**  
**Freeze-Dried: 2°C to 8°C**  
**Live Culture: See Propagation Section**

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Biosafety Level  
**1**

## Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

## Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Chlorobaculum tepidum* (ATCC® 49652™)

American Type Culture Collection  
PO Box 1549  
Manassas, VA 20108 USA  
[www.atcc.org](http://www.atcc.org)

800.638.6597 or 703.365.2700  
Fax: 703.365.2750  
Email: [Tech@atcc.org](mailto:Tech@atcc.org)

Or contact your local distributor

## Description

**Designation:** TLS [CIP 105973]

**Deposited Name:** *Chlorobium tepidum* Wahlund et al.

**Product Description:** Type strain. Genome sequenced strain.

## Propagation

### Medium

ATCC® Medium 37: Chromatium medium

ATCC® Medium 260: Trypticase soy agar/broth with defibrinated sheep blood

### Growth Conditions

**Temperature:** 45°C

**Atmosphere:** Aerobic

### Propagation Procedure

1. Open thawed vial according to enclosed instructions or visit [www.atcc.org](http://www.atcc.org) for instructions.
2. Remove 6 mL of pre-mixed #37 media from primary tube and aseptically transfer the entire contents of thawed vial to that tube. Additional test tubes can be inoculated by transferring up to 2 mL of the primary broth tube to secondary broth tubes. Fill the test tubes to capacity with additional #37 broth, leaving a small air bubble for expansion. Seal the test tube with a screw cap.
3. Incubate at 45°C under a tungsten lamp for 3-7 days. The light should be on a 12 hour cycle (12 hours on/ 12 hours off). Incubate one #260 agar plate aerobically at 37°C to check for contamination.
4. Use 10-20% of an actively growing culture to inoculate fresh tubes of medium.

## Notes

ATCC Medium #37 may be stored separately as solution 1, solution 2/3 and solution 4. In a small screw cap tube, mix 5 mL of solution 1, 3 mL of solution 2/3, 0.2 mL of solution 4, and reduce the media with 0.1 mL of a 10% Thiosulphate solution. Place tubes in the dark for 24 hours prior to inoculation. A hazy, white precipitate should develop in the tubes overnight.

After 3-7 days, growth is evident by turbidity and a deep green pigmentation throughout the broth. When examined microscopically, the cells appear as small short rods in singles and pairs and are non-motile. The culture should be fed sodium sulfide at least once a week. This culture is oxygen-tolerant therefore strictly anoxic conditions are not required when using a large inoculum.

Additional information on this culture is available on the ATCC® web site at [www.atcc.org](http://www.atcc.org).

## References

References and other information relating to this product are available online at [www.atcc.org](http://www.atcc.org).

## Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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## Disclaimers



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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at [www.atcc.org](http://www.atcc.org)

Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).

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