



Product Sheet

Desulfobacula phenolica (ATCC® 43956™)

Please read this FIRST



Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Desulfobacula phenolica* (ATCC® 43956™)

Description

Designation: DSM 3384 [Ph01]

Deposited Name: *Desulfobacterium phenolicum* Bak and Widdel

Propagation

Medium

ATCC® Medium 1628: *Desulfobacterium phenolicum* medium

Growth Conditions

Temperature: 30°C

Atmosphere: Anaerobic gas mixture, 80%N₂ – 20% CO₂

Propagation Procedure

1. Sterilize the top of the Balch tube by spraying it with 70% ethanol and then flame the top.
2. If needed exchange the gas in the test tube for 80% N₂ 20% CO₂.
3. If the medium is pink (see discussion about resazurin) add 2.0 mL of reducing agent (1.5% sodium sulfide, stock solution) per 100 mL of medium. Let the medium sit at room temperature for 10 to 20 minutes until the resazurin becomes colorless before inoculating.
4. When the Balch tube is ready to inoculate, thaw the frozen vial at room temperature under a gentle stream of oxygen-free gas.
5. For inoculation, use a 1.0 mL syringe tipped with 22-gauge needle, withdraw the entire cell suspension from the vial and transfer it to the broth. Using the same needle withdraw 0.5 mL from the broth and inoculate a second Balch tube. Plate 0.1 mL of the inoculated culture onto a non-selective medium and incubate aerobically at 30°C.
6. Growth should be detected in the #1628 broth within 2 to 4 days. There should be no growth detected on the aerobic plate or broth.

NOTE: Addition of 10-20 mg sodium dithionite per liter (e.g. from 5% w/v solution freshly prepared under N₂ and filter-sterilized) may stimulate growth of all strains at the beginning. For transfers use 5-10% inoculum. Incubate all strains in the dark.

ANAEROBIC CONDITIONS:

1. Resazurin is a commonly used redox indicator that is pink when the redox potential is above –50 mv., and colorless when the redox potential is below –110 mv. i.e. highly reducing. Most strict anaerobes require this low redox potential for optimum growth.
2. To obtain a fully reduced medium, it is necessary that the medium be anoxic and that a reducing agent be added. Common reducing agents are sodium sulfide, cysteine, dithiothreitol, and titanium citrate.
3. Syringes can be made anaerobic by one of two methods.
 1. Displace the dead space in the syringe with a sterile oxygen-free gas.
 2. Displace the dead space in the syringe with a reducing agent.

Notes

At 1000x magnification cells are curved rods. The cells stain Gram-negative.

Additional information on this culture is available on the ATCC® web site at www.atcc.org.

References

References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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