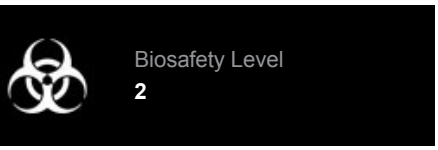
ATCC

Product Sheet

Helicobacter fennelliae (ATCC® 35684™)

Please read this FIRST



Intended Use

This product is intended for research use only. It is not intended for any animal or human therapeutic or diagnostic use.

Citation of Strain

If use of this culture results in a scientific publication, it should be cited in that manuscript in the following manner: *Helicobacter fennelliae* (ATCC® 35684™)

American Type Culture Collection
PO Box 1549
Manassas, VA 20108 USA
www.atcc.org

800.638.6597 or 703.365.2700
Fax: 703.365.2750
Email: Tech@atcc.org

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Description

Designation: 231 [CCUG 18820, LMG 7546]
Deposited Name: *Campylobacter fennelliae* Totten et al.

Propagation

Medium
ATCC® Medium 260: Trypticase soy agar/broth with defibrinated sheep blood

Growth Conditions
Temperature: 37.0°C
Atmosphere: Microaerophilic

Propagation Procedure
1. Open vial according to enclosed instructions. Rehydrate contents of vial with 0.5 ml of Trypticase Soy Broth.
1.
2. To obtain a biphasic culture, add 0.4 ml of the suspension to a #260 slant. Add remaining 0.1 ml of the suspension to a #260 plate and streak for isolation.
3. Incubate at 37ºC under microaerophilic conditions. This organism requires additional free hydrogen for best growth. To obtain this, use an anaerobe jar WITHOUT an active catalyst and an ANAEROBIC gas generator pack, or other acceptable method, to obtain the desired gas mixture. Incubate slant with cap loose.
4. Within 3–5 days of incubation, good growth should be obtained in the broth pool at the bottom of the slant. Additional incubation may be required for colonies to appear on the plate. Further subcultures can be made using broth pool as the inoculum source. Subcultures will require only 24 to 48 hours of incubation.

Notes

This is a slow growing organism that requires moist conditions for best growth. Growth at the broth/agar interface of the biphasic slant should occur within three to five days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy. Organisms are small thin spiral rods. Cells from old cultures may be spherical. The presence of spheroid cells indicates that viability is being lost either due to age or too much exposure to oxygen.
Growth on agar takes longer than with the biphasic culture. Colonies are nonhemolytic, small, gray or clear, and less than l mm in diameter. Spreading may occur with continued incubation.

Once good growth is present, these organisms tend to lose viability, especially if exposed to air for lengthy periods. Viability also decreases with repeated subculturing. Therefore, transfer or freeze the culture when optimal growth is achieved. Adding an equal amount of 20% sterile glycerol to pooled broth from several biphasic slants followed by freezing in liquid nitrogen or *ultra-low temperature* freezer is recommended.

The cells do not Gram stain well using traditional procedures. To obtain the best results, use a basic fuchsin counterstain in place of the safranin.

Additional information on this culture is available on the ATCC web site at www.atcc.org.

References

References and other information relating to this product are available online at www.atcc.org.

Biosafety Level: 2

Appropriate safety procedures should always be used with this material. Laboratory safety is discussed in the current publication of the *Biosafety in Microbiological and Biomedical Laboratories* from the U.S. Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes for Health.

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longer valid.

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Please see the enclosed Material Transfer Agreement (MTA) for further details regarding the use of this product. The MTA is also available on our Web site at www.atcc.org

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