## ATCC Medium 1352: Oxalate Maintenance Agar/Broth Medium

K <sub>2</sub> HPO <sub>4</sub>	0.25 g
KH <sub>2</sub> PO <sub>4</sub>	0.25 g
$(NH_4)_2SO_4$	0.5 g
MgSO <sub>4</sub> x 7H <sub>2</sub> O	0.025 g
Sodium acetate	$0.82 \mathrm{~g}$
Yeast extract	1.0 g
Sodium oxalate	5.0 g
Resazurin	. 0.001 g
Na <sub>2</sub> CO <sub>3</sub>	. 4.0 g
Cysteine HCl x H <sub>2</sub> O	0.5 g
Agar (if required)	. 15.0 g
DI Water	. 1.0 L

*Mix ingredients except Na<sub>2</sub>CO<sub>3</sub> and cysteine.* 

Adjust pH to **6.8**, heat to boiling and then cool while gassing with  $O_2$ -free  $CO_2$ .

Add sodium carbonate and cysteine and tube under a  $CO_2$  atmosphere using Hungate or VPI Anaerobe Lab techniques.

Autoclave in press to hold stopper on tubes. Transfer cultures under CO<sub>2</sub> Atmosphere.

**Note:** For agar medium, prepare and autoclave aerobically. Oxidized Medium browns severely with autoclaving. To avoid this, sterilize first five ingredients in one solution, next three items plus agar in a second solution, sodium carbonate as third solution and filter sterilize cysteine. Combine all aseptically and dispense. Final pH of un-gassed agar is **9.0**. For Slants don't add the sodium carbonate until after boiling.

pH of the medium will drop when incubated in 100% CO<sub>2</sub> atmosphere. Browning does not occur when complete medium is autoclaved anaerobically.