SNU-C2A

CCL-250.1™

Description

SNU-C2A is one of 14 colorectal carcinoma cell lines derived by J.G. Park and associates during the years 1982 through 1985. The cell line is from an Asian, 43-year-old, female patient with colorectal cancer and was obtained from a third passage nude mouse xenograft of tumor fragments and can be used in cancer research.
Organism: *Homo sapiens*, human
Age: 43 years
Gender: Female
Morphology: epithelial
Growth properties: Loosely adherent, multicellular aggregates
Disease: Carcinoma; Colorectal

Storage Conditions

Product format: Frozen
Storage conditions: Vapor phase of liquid nitrogen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to

understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Temperature: 37°C
Atmosphere: 95% Air, 5% CO_2

Handling Procedures

Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the

cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Complete medium: ACL-4 medium supplemented with 10% FBS

The base medium for this cell line is ATCC-formulated DMEM: F12 Medium Catalog No. 30-2006. To make the complete growth medium, add the following components to the base medium:

1. 0.02 mg/ml insulin
2. 0.01 mg/ml transferrin
3. 25 nM sodium selenite
4. 50 nM Hydrocortisone
5. 1 ng/ml Epidermal Growth Factor (do not filter)
6. 0.01 mM ethanolamine
7. 0.01 mM phosphorylethanolamine
8. 100 pM triiodothyronine
9. 0.5% (w/v) bovine serum albumin
10. 10 mM HEPES
11. 0.5 mM sodium pyruvate
12. 2mM L-glutamine(for final conc. of 4.5 mM)
13. 10% FBS

Handling Procedure:

Handling Procedure for Frozen Cells

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at

-70°C. Storage at -70°C will result in loss of viability.

SAFETY PRECAUTION: ATCC highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).

2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

3. Transfer the vial contents to a centrifuge tube containing 9.0 ml complete culture medium and spin at approximately 125 xg for 5 to7 minutes.

4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). and dispense into a new culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

5. Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product

Subculturing procedure:
Protocol: Cultures can be maintained by scraping off the attached cells and transfer along with the the floating cells into new flasks.
Medium Renewal: Two times weekly
Note: Growth is slow.
Reagents for cryopreservation: Complete growth medium supplemented with 10% fetal bovine serum (ATCC 30-2020) and 7.5% (v/v) DMSO (ATCC 4-X)

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: SNU-C2A (ATCC CCL-250.1)

References

References and other information relating to this material are available at www.atcc.org.

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