



Toxoplasma gondii (Nicolle and Manceaux) Nicolle and Manceaux

50174™

Description

Strain designation: RH

Deposited As: *Toxoplasma gondii* (Nicolle and Manceaux) Nicolle and Manceaux

Type strain: No

Genotype: Haplogroup 1

Storage Conditions

Product format: Frozen

Storage conditions: -80°C or colder for 1 week, vapor phase of liquid nitrogen for long-term storage

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 2

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or national agencies.

Product Sheet

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Host:

In vivo cultivation, mouse

Handling Procedures

Storage and Culture Initiation

Frozen ampules packed in dry ice should either be thawed immediately or stored in liquid nitrogen. If liquid nitrogen storage facilities are not available, frozen ampoules may be stored at or below -70°C for approximately one week. **Do not under any circumstance store frozen ampules at refrigerator freezer temperatures (generally**

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-20°C). Storage of frozen material at this temperature will result in the death of the culture.

1. To thaw a frozen ampule, place it in a 35°C water bath such that the lip of the ampule remains above the water line. Thawing time is approximately 2 to 3 minutes. Do not agitate the ampule. Do not leave ampule in water bath after it is thawed.
2. Immediately after thawing, aseptically remove the contents of the ampule with a syringe and inoculate an uninfected mouse. Follow the protocol for maintenance *in vivo*. The course of infection may be longer or shorter than usual depending on percent recovery of the parasite from the frozen state.

Culture maintenance: Tyrod's Salt Solution

NaCl, 8.00 g

KCl, 0.20 g

CaCl₂ 0.20 g

MgCl₂ · H₂O 0.05 g

NaH₂PO₄ · H₂O 1.00 g

NaHCO₃ · H₂O 1.00 g

Glucose 1.00 g

Glass distilled H₂O to 1.00 L

Add ingredients in the sequence listed. Filter-sterilize.

1. Inject the entire contents of the thawed ampule intraperitoneally into a 6- to 9-week-old mouse. The infection should be well developed within 6-9 days post inoculation. The abdomen of the mouse will become increasingly swollen as the infection progresses.
2. When the infection is well developed, remove the peritoneal fluid from the mouse using the following technique:
 - a. Inject 2 mL of Tyrod's solution into the peritoneum; massage the stomach for 2 minutes, and then using a 5-mL syringe, remove the peritoneal fluid (~3 mL).
 - b. Inoculate 0.2 mL of this fluid per mouse to subculture.

Cryopreservation:

1. Harvest the parasites according to the protocol for maintenance *in vivo*.
2. Spin the cell suspension at approximately 50 x g for 3 min, to remove the cellular debris.
3. Transfer the supernatant to a new 15 mL plastic centrifuge tube. Centrifuge at 1300 x g for 10 min.
4. Pool the cell pellets and adjust the concentration to 2.0 - 4.0 x 10⁷ cells/mL

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with a fresh solution of Tyrode's Salt Solution.

*If the concentration is too low centrifuge at 1300 x g for 10 min and resuspend in the volume of Tyrode's Salt Solution required to yield the desired concentration.

5. Mix the cell preparation and 15% (v/v) DMSO in equal portions. The final concentration will be $1.0 - 2.0 \times 10^7$ cells/mL and 7.5% DMSO. The time from the mixing of the cell preparation and cryoprotective solution to the start of the freezing process should be no less than 15 min. and no more than 30 min.
6. Dispense in 0.5 mL aliquots to 1.0-2.0 mL sterile plastic screw-capped cryules (special plastic vials for cryopreservation).
7. Place the vials in a controlled rate freezing unit. From room temperature cool at $-1^\circ\text{C}/\text{min}$ to -40°C . If the freezing unit can compensate for the heat of fusion, maintain rate at $-1^\circ\text{C}/\text{min}$ through the heat of fusion. At -40°C plunge into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately $-1^\circ\text{C}/\text{min}$.)
8. Store in either the vapor or liquid phase of a nitrogen refrigerator.
9. To thaw a frozen ampule, place it in a 35°C water bath such that the lip of the ampule remains above the water line. Thawing time is approximately 2 to 3 minutes. Do not agitate the ampule. Do not leave ampule in water bath after thawed.
10. Immediately after thawing, aseptically remove the contents of the ampule with a syringe and inoculate an uninfected mouse. Follow the protocol for maintenance *in vivo*.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Toxoplasma gondii* (Nicolle and Manceaux) Nicolle and Manceaux (ATCC 50174)

References

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References and other information relating to this material are available at www.atcc.org.

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Product Sheet

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Contact Information

ATCC

10801 University Boulevard

Manassas, VA 20110-2209

USA

***Toxoplasma gondii* (Nicolle and Manceaux) Nicolle and Manceaux**

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US telephone: 800-638-6597

Worldwide telephone: +1-703-365-2700

Email: tech@atcc.org or contact your local distributor
