



# ***Oxobacter pfennigii*** **(Krumholz and Bryant)** **Collins et al.**

**43583™**

## **Description**

**Strain designation:** DSM 3222 [V5-2]

**Deposited As:** *Clostridium pfennigii* Krumholz and Bryant

**Type strain:** Yes

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## **Storage Conditions**

**Product format:** Freeze-dried

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## **Intended Use**

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## **BSL 1**

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## Growth Conditions

**Medium:**

ATCC Medium 1644: *Clostridium pfennigii* medium

**Temperature:** 37°C

**Atmosphere:** Anaerobic

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## Handling Procedures

1. Open vial according to enclosed instructions.
2. Under anaerobic conditions, withdraw 0.5 ml of #1644 from a single test tube (5 to 6 ml) and rehydrate the vial contents.
3. Aseptically transfer this aliquot back into the broth tube. Additional tubes may be inoculated with 0.5 ml each from the suspension. A slant of #1644 may also be

**inoculated with 0.2 ml. Streak several blood plates to check for colonial morphology and purity.**

**4. Incubate tubes under an anaerobic atmosphere at 37°C. Incubate one agar plate anaerobically for colony formation, and one aerobically for aerobic contamination check.**

#### ANAEROBIC CONDITIONS:

Anaerobic conditions for transfer may be obtained by either of the following:

- Use of an anaerobic gas chamber, or
- Placement of test tubes under a gassing cannula system hooked to anaerobic gas.

Anaerobic conditions for incubation may be obtained by any of the following:

- Loose screw caps on test tubes in anaerobic chamber,
- Loose screw caps on test tubes in an activated anaerobic gas pack jar, or
- Use of sterile butyl rubber stoppers on test tubes so that an anaerobic gas headspace is retained.

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## Notes

Within 48-72 hours, growth is evident by slight turbidity in the broth. No growth should occur on the blood agar plate incubated aerobically.

Additional information on this culture is available on the ATCC web site at [www.atcc.org](http://www.atcc.org).

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## Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Oxobacter pfennigii* (Krumholz and Bryant) Collins et al. (ATCC 43583)

## References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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## Contact Information

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**43583**

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