



# Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011<sup>TM</sup>

## Description

Primary Bronchial Tracheal Smooth Muscle Cells are myocytes isolated from the respiratory tract of a donor and can be used in the study of angiogenesis, atherosclerosis, diabetes, or vascular/pulmonary biology.

**Organism:** *Homo sapiens*, human

**Cell Type:** myocyte

**Tissue:** Respiratory tract

**Age:** lot-specific

**Gender:** Lot-specific

**Morphology:** elongated, spindle-shaped

**Growth properties:** Adherent

**Disease:** Normal

**Volume:** 1.0 mL

---

## Storage Conditions

**Product format:** Frozen

**Storage conditions:** Vapor phase of liquid nitrogen

---

## Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

---

# Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011

**BSL 1**

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

All tissues used for isolation are obtained under informed consent and conform to HIPAA regulations to protect the privacy of the donor's Personally Identifiable Information. It is best to use caution when handling any human cells. We recommend that all human cells be accorded the same level of biosafety consideration as cells known to carry Human immunodeficiency virus (HIV) and other bloodborne pathogens. With infectious virus assays or viral antigen assays, even a negative test result may not exclude the possibility of the existence of a latent viral genome or infectious viral particles below the lower limit of detection of that assay.

ATCC recommends that appropriate safety procedures be used when handling all primary cells and cell lines, especially those derived from human or other primate material. Handle as a potentially biohazardous material using universal precautions. Cells derived from primate lymphoid tissue may fall under the regulations of 29 CFR 1910.1030 Bloodborne Pathogens.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

# Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011

## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

## Handling Procedures

### Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

**Complete medium:** 1. Obtain one growth kit from the freezer; make sure that the caps of all components are tight.

2. Thaw the components of the growth kit just prior to adding them to the basal medium. It is necessary to warm the L-glutamine component in a 37°C water bath and shake to dissolve any precipitates prior to adding to the basal medium.
3. Obtain one bottle of Vascular Cell Basal Medium (475 mL) from cold storage.
4. Decontaminate the external surfaces of all growth kit component vials and the basal medium bottle by spraying them with 70% ethanol.
5. Using aseptic technique, and working in a laminar flow hood or biosafety cabinet, transfer the volume of each growth kit component, as indicated below, to the bottle of basal medium using a separate sterile pipette for each transfer.
6. Tightly cap the bottle of complete growth medium and swirl the contents gently to assure a homogeneous solution. Do not shake forcefully to avoid foaming. Label and date the bottle.
7. Complete growth media should be stored in the dark at 2°C to 8°C (do not freeze). When stored under these conditions, complete growth media is stable for 30 days.

### Vascular Smooth Muscle Cell Growth Kit Components:

rh FGF-basic, 0.5 mL, 5 ng/mL  
rh Insulin, 0.5 mL, 5 µg/mL

## Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

**PCS-130-011**

Ascorbic acid, 0.5 mL, 50 µg/mL  
L-glutamine, 25.0 mL, 10 mM  
rh EGF, 0.5 mL, 5 ng/mL  
Fetal Bovine Serum, 25.0 mL, 5%

Antimicrobials and phenol red are not required for proliferation but may be added if desired. The recommended volume of either of the optional components (GA solution or PSA solution) to be added to the complete growth media is summarized below

### Addition of Antimicrobials/Antimycotics and Phenol Red (Optional):

Gentamicin-Amphotericin B Solution, 0.5 mL, (Gentamicin: 10 µg/mL, Amphotericin B: 0.25 µg/mL)

Penicillin-Streptomycin-Amphotericin B Solution, 0.5 mL, (Penicillin: 10 Units/mL, Streptomycin: 10 µg/mL, Amphotericin B: 25 ng/mL)

Phenol Red, 0.5 mL, 33 µM

### Reagents for subculture::

1. DPBS (ATCC 30-2200)
2. Trypsin-EDTA for Primary Cells (ATCC PCS-999-003) containing 0.05% Trypsin and 0.02% EDTA.
3. Trypsin Neutralizing Solution (ATCC PCS-999-004)

**Required media:** One bottle of ATCC PCS-100-030 Vascular cell Basal Medium (475mL) plus one of the Vascular Smooth Muscle Cell Growth Kit (ATCC PCS-100-042) that includes the following growth components: rh FGF-basic, rh Insulin, Ascorbic acid, L-Glutamine, rh EGF, Fetal Bovine Serum

### Optional media supplements:

1. Gentamicin-Amphotericin B Solution (ATCC PCS-999-025)
2. Penicillin-Streptomycin-Amphotericin B Solution (ATCC PCS-999-002)
3. Phenol Red (ATCC PCS-999-001)

### Handling Procedure:

1. ***Refer to the batch specific information for the total number of viable cells recovered from this lot of ATCC® PCS-130-011.***
2. Using the total number of viable cells, determine how much surface area can be inoculated to achieve an initial seeding density of 5,000 cells per cm<sup>2</sup>.
3. Prepare the desired combination of flasks. Add 5 mL of complete growth media per 25 cm<sup>2</sup> of surface area. Place the flasks in a 37°C, 5% CO<sub>2</sub>, humidified incubator and allow the media to pre-equilibrate to temperature and pH for 30

## Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011

minutes prior to adding cells.

4. While the culture flasks equilibrate, remove one vial of ATCC® PCS-130-011 from storage and thaw the cells by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 1 to 2 minutes).
5. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. *All operations from this point onward should be carried out under strict aseptic conditions.*
6. Add the appropriate volume of complete growth media [volume = (1 mL x number of flasks to be seeded) - 1 mL] into a sterile conical tube. Using a sterile pipette, transfer the cells from the cryovial to the conical tube. Gently pipette the cells to homogenize the suspension. Centrifugation may be performed to remove DMSO.
7. Transfer 1.0 mL of the cell suspension to each of the pre-equilibrated culture flasks prepared in steps 1 to 3 of *Handling Procedure for Frozen Cells and Initiation of Culture*. Pipette several times, then cap and gently rock each flask to evenly distribute the cells.
8. Place the seeded culture flasks in the incubator at 37°C with a 5% CO<sub>2</sub> atmosphere. Incubate for at least 24 hours before processing the cells further.

### Subculturing procedure:

1. Passage normal bronchial tracheal smooth muscle cells when culture has reached approximately 80% confluence.
2. Warm both the Trypsin-EDTA for Primary Cells (ATCC PCS-999-003) and the Trypsin Neutralizing Solution (ATCC PCS-999-004) to room temperature prior to dissociation. Warm complete growth medium to 37°C prior to use with the cells.
3. For each flask, carefully aspirate the spent media without disturbing the monolayer.
4. Rinse the cell layer two times with 3 to 5 mL D-PBS (ATCC 30-2200) to remove residual traces of serum.
5. Add pre-warmed trypsin-EDTA solution (1 to 2 mL for every 25 cm<sup>2</sup>) to each flask.
6. Gently rock each flask to ensure complete coverage of the trypsin-EDTA solution over the cells, and then aspirate the excess fluid off of the monolayer.
7. Observe the cells under the microscope. When the cells pull away from each other and round up (typically within 3 to 5 minutes), remove the flask from the microscope and gently tap it from several sides to promote detachment of the



ATCC® Credible leads to Incredile®

[www.atcc.org](http://www.atcc.org)

# Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011

cells from the flask surface.

8. When the majority of cells appear to have detached, quickly add an equal volume of Trypsin Neutralizing Solution (ATCC PCS-999-004) to each flask. Gently pipette or swirl the culture to ensure all of the trypsin-EDTA solution has been neutralized.
9. Transfer the dissociated cells to a sterile centrifuge tube and set aside while processing any remaining cells in the flask.
10. Add 3 to 5 mL D-PBS (ATCC 30-2200) to the flask to collect any additional cells that might have been left behind.
11. Transfer the cell/D-PBS suspension to the centrifuge tube containing the trypsin-EDTA-dissociated cells.
12. Repeat steps 10 and 11 as needed until all cells have been collected from the flask.
13. Centrifuge the cells at 150 x g for 3 to 5 minutes.
14. Aspirate the neutralized dissociation solution from the cell pellet and resuspend the cells in 2 to 8 mL fresh, pre-warmed, complete growth medium.
15. Count the cells and seed new flasks at a density of 5,000 cells per cm<sup>2</sup>.
16. Place newly seeded flasks in a 37°C, 5% CO<sub>2</sub>, incubator for at least 24 to 48 hours before processing the cells further. Refer to *Maintenance* for guidelines on feeding.

## Culture maintenance:

1. Before beginning, pre-warm complete growth media in a 37°C water bath. This will take between 10 and 30 minutes, depending on the volume. If using a small volume of medium (50 mL or less), warm only the volume needed in a sterile conical tube. **Avoid warming complete growth media multiple times.**
2. 24 to 36 hours after seeding, remove the cells from the incubator and view each flask under the microscope to determine percent cellular confluence.
3. Carefully remove the spent media without disturbing the monolayer.
4. Add 5 mL of fresh, pre-warmed complete growth media per 25 cm<sup>2</sup> of surface area and return the flasks to the incubator.
5. After 24 to 48 hours, view each flask under the microscope to determine percent cellular confluence. If not ready to passage, repeat steps 3 and 4 as described above. When cultures have reached approximately 80% confluence, and are actively proliferating (many mitotic figures are visible), it is time to subculture. Bronchial Tracheal smooth muscle cells are contact inhibited; post-confluent bronchial tracheal smooth muscle cells may not proliferate after passaging.



# Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011

**Note:** Cells are typically ready to passage after 4 to 6 days in culture when inoculated with 5,000 cells/cm<sup>2</sup>.

---

## Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human (ATCC PCS-130-011)

---

## References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

---

## Warranty

The product is provided 'AS IS' and the viability of ATCC<sup>®</sup> products is warranted for 30 days from the date of shipment, provided that the customer has stored and handled the product according to the information included on the product information sheet, website, and Certificate of Analysis. For living cultures, ATCC lists the media formulation and reagents that have been found to be effective for the product. While other unspecified media and reagents may also produce satisfactory results, a change in the ATCC and/or depositor-recommended protocols may affect the recovery, growth, and/or function of the product. If an alternative medium formulation or reagent is used, the ATCC warranty for viability is no longer valid. Except as expressly set forth herein, no other warranties of any kind are provided, express or implied, including, but not limited to, any implied warranties of merchantability, fitness for a particular purpose, manufacture according to cGMP standards, typicality, safety, accuracy, and/or noninfringement.

---

# Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011

## Disclaimers

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use. Any proposed commercial use is prohibited without a [license from ATCC](#).

While ATCC uses reasonable efforts to include accurate and up-to-date information on this product sheet, ATCC makes no warranties or representations as to its accuracy. Citations from scientific literature and patents are provided for informational purposes only. ATCC does not warrant that such information has been confirmed to be accurate or complete and the customer bears the sole responsibility of confirming the accuracy and completeness of any such information.

This product is sent on the condition that the customer is responsible for and assumes all risk and responsibility in connection with the receipt, handling, storage, disposal, and use of the ATCC product including without limitation taking all appropriate safety and handling precautions to minimize health or environmental risk. As a condition of receiving the material, the customer agrees that any activity undertaken with the ATCC product and any progeny or modifications will be conducted in compliance with all applicable laws, regulations, and guidelines. This product is provided 'AS IS' with no representations or warranties whatsoever except as expressly set forth herein and in no event shall ATCC, its parents, subsidiaries, directors, officers, agents, employees, assigns, successors, and affiliates be liable for indirect, special, incidental, or consequential damages of any kind in connection with or arising out of the customer's use of the product. While reasonable effort is made to ensure authenticity and reliability of materials on deposit, ATCC is not liable for damages arising from the misidentification or misrepresentation of such materials.

Please see the material transfer agreement (MTA) for further details regarding the use of this product. The MTA is available at [www.atcc.org](http://www.atcc.org).

---

## Copyright and Trademark Information

© ATCC 2023. All rights reserved.

ATCC is a registered trademark of the American Type Culture Collection.

---

# Primary Bronchial/Tracheal Smooth Muscle Cells; Normal, Human

PCS-130-011

Revision

This information on this document was last updated on 2025-12-05

---

## Contact Information

ATCC

10801 University Boulevard

Manassas, VA 20110-2209

USA

US telephone: 800-638-6597

Worldwide telephone: +1-703-365-2700

Email: [tech@atcc.org](mailto:tech@atcc.org) or contact your local distributor

---



ATCC® Credible leads to Incredible®

[www.atcc.org](http://www.atcc.org)