



MeT-5A

CRL-9444™

Description

MeT-5A is an epithelial cell line from the mesothelium that was isolated from the pleural fluids of non-cancerous individuals. The cells were transfected with the pRSV-T plasmid (an SV40 or-construct containing the SV40 early region and the Rous sarcoma virus long terminal repeat) and cloned.

Organism: *Homo sapiens*, human

Cell Type: epithelial cell

Tissue: Mesothelium

Morphology: epithelial

Growth properties: Adherent

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Storage Conditions

Product format: Frozen

Storage conditions: Vapor phase of liquid nitrogen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 2

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Cells contain polyomaviral DNA sequences

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Temperature: 37°C

Handling Procedures

Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Complete medium: The base medium for this cell line is Medium 199 containing 1.5 g/L sodium bicarbonate.

To make the complete growth medium, add the following components to the base medium:

10% fetal bovine serum (final conc.)

3.3 nM epidermal growth factor (EGF) (final conc.) (do not filter).

400 nM hydrocortisone (final conc.)

870 nM zinc-free bovine insulin (final conc.)

20 mM HEPES (final conc.)

The trace elements at the following final concentrations:

H₂SeO₃ 0.3869 mg/L (Selenious acid)

MnCl₂×4H₂O 0.0198 mg/L (Manganese chloride)

Na₂SiO₃×9H₂O 14.2100 mg/L (Sodium silicate)

(NH₄)₆Mo₇O₂₄×4H₂O 0.1236 mg/L (Ammonium molybdate)

NH₄VO₃ 0.0585 mg/L (Ammonium vanadate)

NiSO₄×6H₂O 0.0131 mg/L (Nickel sulfate)

SnCl₂×2H₂O 0.0113 mg/L (Tin Chloride)

See Methods in Cell Biology, Vol. 21B, pg. 200, 1980

This medium is formulated for use with a 5% CO₂ in air atmosphere.

ATCC tested fetal bovine serum is available as ATCC Catalog No. 30-2020 (500ml) and

ATCC Catalog No. 30-2021 (100ml).

Handling Procedure:

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. It is recommended that the cryoprotective agent be removed immediately. Centrifuge the cell suspension at approximately 125 x g for 5 to 10 minutes. Discard the supernatant and resuspend the cell pellet in an appropriate amount of fresh growth medium.
4. Transfer the vial contents to an appropriate size vessel. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Subculturing procedure: Volumes are given for a 75 cm² flask. Increase or decrease the amount of dissociation medium needed proportionally for culture vessels of other sizes.

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin- 0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently

pipetting.

5. To remove trypsin-EDTA solution, transfer cell suspension to centrifuge tube and spin at approximately 125 x g for 5 to 10 minutes. Discard supernatant and resuspend cells in fresh growth medium. Add appropriate aliquots of cell suspension to new culture vessels.

6. Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 to 1:4 is recommended

Medium Renewal: Every 2 to 3 days

Reagents for cryopreservation: Complete growth medium supplemented with 5% (v/v) DMSO (ATCC 4-X)

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: MeT-5A (ATCC CRL-9444)

References

References and other information relating to this material are available at www.atcc.org.

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