



HBEC3-KT

CRL-4051™

Product Sheet

Description

HBEC3-KT is an hTERT-immortalized epithelial cell that was isolated from the bronchus of a 65-year-old, female patient. This cell line was deposited by John Minna and Shelley Sheridan, and can be used in toxicology research.

Cell Type: epithelial cell

Tissue: Lung; Bronchus

Age: 65 years

Gender: Female

Morphology: epithelial, packed cuboidal

Growth properties: Adherent

Disease: Normal

Storage Conditions

Product format: Frozen

Storage conditions: Vapor phase of liquid nitrogen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 2

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to

understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

Cells immortalized by CDK5

Cells immortalized by hTERT

Cells contain SV40 promoter sequences

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Temperature: 37°C

Atmosphere: 95% Air, 5% CO₂

Handling Procedures

Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C , preferably in liquid nitrogen vapor, until ready for use.

Complete medium: Airway Epithelial Cell Basal Medium (ATCC PCS-300-030) supplemented with Bronchial Epithelial Cell Growth Kit (ATCC PCS-300-040)

Handling Procedure: To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt.

If storage of the frozen culture is necessary upon arrival, store the vial in liquid nitrogen vapor phase and NOT

at -70°C . Storage at -70°C will result in loss of viability.

1. Prepare a 25 cm^2 or a 75 cm^2 culture flask containing the recommended complete culture medium. Prior to the addition of the vial contents, the vessel containing the growth medium should be placed in the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6) and to avoid excessive alkalinity of the medium during recovery of the cells.
2. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
3. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All operations from this point on should be carried out under strict aseptic conditions.
4. Transfer the vial contents to a centrifuge tube containing 3.0 mL of complete culture medium, mix gently, then slowly add an additional 7.0 mL of complete growth medium and centrifuge the cell suspension at approximately 1000 rpm for 5 minutes at room temperature.
5. Discard the supernatant (ENSURING THAT AS MUCH OF THE FBS/DMSO IS REMOVED AS THESE CELLS ARE SENSITIVE TO IT) and resuspend the cells in 3 mL of fresh growth medium. Count the cells and seed at recommended seeding density.
6. Incubate the culture at 37°C in a suitable incubator.

Subculturing procedure: Volumes used in this protocol are for 75 cm^2 flasks; proportionally reduce or increase amount of dissociation solutions for culture vessels of other sizes.

Subculture when the culture is about 70-80% confluent.

1. Remove and discard spent medium.
2. Briefly rinse the cells with Dulbecco's Phosphate Buffered Saline (DPBS, ATCC 30-2200), 1 mL / 25 cm² and discard rinse solution.
3. Add Trypsin-EDTA, at 1 mL / 25 cm², for Primary Cells (ATCC PCS-999-003) to the flask. Incubate at 37°C for 4-6 min (until 90% of the cells have detached).
4. tap flask gently to ensure cells are detached. Add 2% FBS in DPBS at 1 mL / 25 cm² to neutralize the trypsin.
5. Centrifuge cells at 1000rpm for 5 min at room temperature.
6. Remove supernatant. Resuspend pellet in 6.0 to 8.0 mL Complete Growth Medium.
7. Count cells, and seed 4.0 x 10³ to 6.0 x 10³ viable cells/cm² to new culture vessels.

Medium Renewal: Every 2-3 days.

Note: The cells are sensitive to DMSO and FBS. Ensure that as much as possible is removed after centrifugation, and before seeding into the recommended culture medium.

Reagents for cryopreservation: Complete growth medium supplemented with 10% (v/v) FBS and 10% (v/v) DMSO (ATCC 4-X).

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: HBEC3-KT (ATCC CRL-4051)

References

References and other information relating to this material are available at www.atcc.org.

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Revision

This information on this document was last updated on 2025-01-06

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