



# BJ-5ta

CRL-4001™

## Description

BJ-5ta is an hTERT-immortalized fibroblast cell that was isolated from the foreskin of a male patient. This cell line was deposited by Geron Corporation and can be used in toxicology research.

**Organism:** *Homo sapiens*, human

**Cell Type:** fibroblast

**Tissue:** Skin; Foreskin

**Age:** neonate

**Gender:** Male

**Morphology:** Fibroblast-like

**Growth properties:** Adherent

**Disease:** Normal

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## Storage Conditions

**Product format:** Frozen

**Storage conditions:** Vapor phase of liquid nitrogen

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## Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories*

(BMBL), U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## Growth Conditions

**Temperature:** 37°C

**Atmosphere:** 95% Air, 5% CO<sub>2</sub>

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## Handling Procedures

### Unpacking and storage instructions:

1. Check all containers for leakage or breakage.

2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below  $-130^{\circ}\text{C}$ , preferably in liquid nitrogen vapor, until ready for use.

**Complete medium:** A 4:1 mixture of Dulbecco's medium and Medium 199 with supplements as follows :

4 parts of Dulbecco's Modified Eagle's Medium containing 4 mM L-glutamine, 4.5 g/L glucose and 1.5 g/L sodium bicarbonate

1 part of Medium 199

Supplemented with:

0.01 mg/ml hygromycin B

10% fetal bovine serum

**Handling Procedure:** To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If storage of the frozen culture is necessary upon arrival, store the vial in liquid nitrogen vapor phase and NOT at  $-70^{\circ}\text{C}$ . Storage at  $-70^{\circ}\text{C}$  will result in loss of viability.

1. Prepare a 25 cm<sup>2</sup> or a 75 cm<sup>2</sup> culture flask containing the recommended complete culture medium. Prior to the addition of the vial contents, the vessel containing the growth medium should be placed in the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6) and to avoid excessive alkalinity of the medium during recovery of the cells.
2. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
3. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All operations from this point on should be carried out under strict aseptic conditions.
4. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge the cell suspension at approximately 125 x g for 5 to 7 minutes.
5. Discard the supernatant and resuspend the cells in fresh growth medium (see the batch-specific information for the recommended dilution ratio). Add this suspension to the prepared culture vessel.
6. Incubate the culture at 37°C in a suitable incubator.
7. A 5% CO<sub>2</sub>/95% air atmosphere is recommended if using the medium described on this product sheet.

**Subculturing procedure:**

Volumes are given for a 75 cm<sup>2</sup> flask. Recommended use of Corning® T-75 flasks

(catalog #430641). Increase or decrease the amount of dissociation medium needed proportionally for culture vessels of other sizes.

1. Remove and discard culture medium.
2. Add 3.0 to 5.0 mL of 0.25% trypsin-0.53 mM EDTA solution to the flask and observe cells under an inverted microscope until the cell layer is dispersed (usually within 5 to 15 minutes).

**Note:** To avoid clumping do not hit or shake the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

3. Add 6.0 to 10.0 mL of complete growth medium and aspirate cells by gently pipetting.
4. Add appropriate aliquots of the cell suspension to new culture vessels. An inoculum of  $3 \times 10^3$  to  $5 \times 10^3$  viable cells/cm<sup>2</sup> is recommended. Maintain cultures at a cell concentration between  $8 \times 10^3$  and  $1 \times 10^4$  cells/cm<sup>2</sup>.
5. Subcultivation ratio: 1:2 to 1:3 twice weekly
6. Incubate cultures at 37°C.

**Subcultivation Ratio:** 1:2 to 1:3 twice weekly

**Medium Renewal:** every 2 to 3 days

**Note:** Subculture when cell concentration reaches between  $8 \times 10^3$  and  $1 \times 10^4$  cells/cm<sup>2</sup>.

**Note:** For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 13 in *Culture Of Animal Cells: A Manual of Basic Technique* by R. Ian Freshney, 5th edition, published by Wiley-Liss, N.Y., 2005.

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## Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: BJ-5ta (ATCC CRL-4001)

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## References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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