



Schneider's Drosophila Line 2 [D. Mel. (2), SL2]

CRL-1963™

Description

Organism: *Drosophila melanogaster*, fruit fly

Tissue: Embryo

Morphology: epithelial

Growth properties: Mixed: suspension with some loosely adherent cells

Patent number: Schneider's Drosophila Line 2 [D. Mel. (2), SL2] is a cell line exhibiting epithelial-like morphology that was isolated in 1969 from the embryo of a fruit fly. This cell line was deposited IR Schneider.

Storage Conditions

Product format: Frozen

Storage conditions: Vapor phase of liquid nitrogen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies

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ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Temperature: 23°C (22-24°C)

Handling Procedures

Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Complete medium:

Schneider's Drosophila Medium, 90%; heat-inactivated fetal bovine serum, 10%.

NOTE: the fetal bovine serum should be selected for ability to support the growth of insect cell lines.

This medium is formulated for use without CO₂.

Example media preparation:

- 500 mL Schneider's Drosophila Medium (Gibco catalog # 21720-024)
- 56 mL heat-inactivated, insect tested FBS (Sigma catalog #F4135)

Handling Procedure:

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C.

Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 24°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a 25 cm² non-vented tissue culture flask and dilute with the recommended complete culture medium (see the lot information on Certificate of Analysis (COA) for the culture recommended dilution ratio).

The recommended seeding density for CRL-1963 is 2.0×10^5 to 4.0×10^5 viable cells/mL.

4. Incubate the culture at 24°C in a suitable incubator.

If it is desired that the cryoprotective agent be removed immediately, or that a more concentrated cell suspension be obtained, centrifuge the cell suspension at approximately 280 x *g* for 10 minutes. Discard the supernatant and resuspend the cells with fresh growth medium at the dilution ratio recommended in the specific batch information.

Note: Cells grow mainly in suspension with minimal loosely attached cells. Treat cell line as a suspension cell line when expanding these cells.

Subculturing procedure:

Gently resuspend the cells by pipetting medium across the monolayer or (for larger flasks) by slapping the flask against the base of the palm. **Note:** Attached cells may be removed by gently banging or tapping the vessels, or by rinsing the growth surface with culture medium. Dilute the cell suspension to the desired concentration with fresh medium. Cultures should be maintained by the addition of fresh medium.

Maintain cultures at a cell concentration between 5×10^4 and 4×10^5 cells/mL.

Subcultivation Ratio: A subcultivation ratio of 1:10 or greater is recommended

Medium Renewal: At the time of subcultivation

Note: The cells will grow as a loose monolayer (or in suspension) at any temperature between 16°C and 28°C.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: Schneider's Drosophila Line 2 [D. Mel. (2), SL2] (ATCC CRL-1963)

References

References and other information relating to this material are available at www.atcc.org.

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