F9

CRL-1720™

Description

F9 is an epithelial-like cell that was isolated from the testis of a mouse with embryonal testicular teratoma and can be used in cancer research. This cell line was deposited by S Strickland.

Organism: *Mus musculus*, mouse
Tissue: Testis
Age: embryo
Morphology: epithelial
Growth properties: Adherent
Disease: Carcinoma; Embryonal testicular teratoma

Storage Conditions

Product format: Frozen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies

and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Temperature: 37°C

Handling Procedures

Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Complete medium:

The base medium for this cell line is ATCC-formulated Dulbecco's Modified Eagle's Medium, Catalog No. 30-2002. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.

Example media preparation:

- 500 mL DMEM (ATCC 30-2002)
- 56 mL FBS (ATCC 30-2020). Not heat-inactivated

Handling Procedure:

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 280 x *g* for 10 minutes.
4. Resuspend cell pellet with the recommended complete growth medium (see the lot information on Certificate of Analysis (COA) for the culture recommended dilution ratio) and transfer into a 25 cm^2 or a 75 cm^2 culture flask coated with 0.1% gelatin coating (see coating instructions in the subculturing procedure section)

   The recommended seeding density for CRL-1720 is 4.0 x 10^4 to 6.0 x 10^4 viable cells/cm^2.

   It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the

incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

5. Incubate the culture at 37°C in a suitable incubator. A 5% CO2 in air atmosphere is recommended if using the medium described on this product sheet.

Subculturing procedure:

Volumes are given for a 75 cm^2 flask. Increase or decrease the amount of dissociation medium needed proportionally for culture vessels of other sizes. NOTE: culture vessels must be coated with 0.1% gelatin prior to use. To do so, cover the surface of the vessel with 0.1% Gelatin Solution (ATCC PCS-999-027) and rock at slow speed for at least 30 minutes, and up to overnight, at 37°C. Wash with DPBS or culture medium prior to use. It is recommended to coat the vessel fresh immediately prior to use; however coated vessel(s) can be stored at 2 - 8°C with coating solution under sterile conditions using a plug seal cap or parafilm.

Note: Culture flasks must be coated with 0.1% gelatin prior to use. To do so, cover the surface of the vessel with 0.1% Gelatin Solution (ATCC PCS-999-027) and rock at slow speed for at least 30 minutes, and up to overnight, at 37°C. Wash with DPBS or culture medium prior to use. Remove rinsing solution and pre-warm the vessel(s) with culture medium at 37C for 1 hour prior to seeding cells. It is recommended to coat the vessel fresh immediately prior to use; however coated vessel(s) can be stored at 2 - 8°C with coating solution under sterile conditions using a plug seal cap or parafilm.

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin- 0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).

   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new coated culture vessels.
6. Incubate cultures at 37°C.

Subcultivation Ratio/Subculture seeding density: A subcultivation ratio of 1:10 is

recommended or subculture at a seeding density of 2.0 x 10^4 to 6.0 x 10^4 viable cells/cm^2 is recommended.
Note: Unless floating cells have been verified to be non-viable via a trypan blue or some other sort of viability assay, floating cells should be kept with the main culture.
Medium Renewal: Every 2 to 3 days

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: F9 (ATCC CRL-1720)

References

References and other information relating to this material are available at www.atcc.org.

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