



# *Tindallia californiensis* Pikuta et al.

**BAA-393™**

## Description

**Strain designation:** APO [CIP 107910, DSM 14871]

**Deposited As:** *Tindallia californica*

**Type strain:** Yes

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## Storage Conditions

**Product format:** Freeze-dried

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## Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always

used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## Growth Conditions

**Medium:**

ATCC Medium 2310: Medium for *Tindallia californica*

**Temperature:** 37°C**Atmosphere:** Anaerobic

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## Handling Procedures

1. Sterilize the top of the Balch tube by spraying it with 70% ethanol and then flame the top.
2. If needed exchange the gas in the test tube for 80% N<sub>2</sub> 20% CO<sub>2</sub>.
4. When the Balch tube is ready to inoculate, thaw the frozen vial at room temperature under a gentle stream of oxygen-free gas.

5. For inoculation, use a 1.0 ml syringe tipped with 22-gauge needle, withdraw the cell suspension from the vial and transfer it to the broth. Plate 0.1 ml of the inoculated culture onto a non-selective medium and incubate aerobically at 37°C. Use 0.1 ml of the inoculated culture to inoculate a nonselective aerobic broth. Incubate the inoculated tubes at 37°C.

6. Growth should be detected in the #2310 broth within 24 to 48 hours. There should be no growth detected on the aerobic plate or broth.

#### ANAEROBIC CONDITIONS:

a. Resazurin is a commonly used redox indicator that is pink when the redox potential is above 50 mv., and colorless when the redox potential is below 110 mv. i.e. highly reducing. Most strict anaerobes require this low redox potential for optimum growth.

b. To obtain a fully reduced medium, it is necessary that the medium be anoxic and that a reducing agent be added. Common reducing agents are sodium sulfide, cysteine, dithiothreitol, and titanium citrate.

c. Syringes can be made anaerobic by one of two methods. 1. Displace the dead space in the syringe with a sterile

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### Notes

At 1000x magnification cells are straight to curved motile rods with rounded edges. The cells stain Gram-positive. Optimal pH is 9.5 with a pH range of 8.0 to 11.0.

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### Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Tindallia californiensis* Pikuta et al. (ATCC BAA-393)

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### References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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## Revision

This information on this document was last updated on 2024-10-25

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## Contact Information

ATCC

10801 University Boulevard

Manassas, VA 20110-2209

USA

## ***Tindallia californiensis* Pikuta et al.**

**BAA-393**

US telephone: 800-638-6597

Worldwide telephone: +1-703-365-2700

Email: [tech@atcc.org](mailto:tech@atcc.org) or contact your local distributor

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