

Desulfovibrio sp.

B∆∆-1916[™]

Description

Strain designation: G11 [OCM 18]

Type strain: No

Storage Conditions

Product format: Frozen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL₁

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories* (*BMBL*), U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and



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will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Handling Procedures

- 1. Sterilize the top of the tube by spraying it with 70% ethanol and then flaming the top.
- 2. If needed, exchange the gas in the test tube for 80% N₂ 20% CO₂.
- 3. Reduce the medium with the addition of reducing agent (see note for Component IV and notes under reducing agent). Let the medium sit at room temperature for at least one hour before inoculation.
- 4. When the Balch tube is ready to inoculate, thaw the frozen vial at room temperature under a gentle stream of oxygen-free gas.
- 5. For inoculation, using an anaerobic 1.0 ml syringe tipped with sterile needle, withdraw the cell suspension from the vial and transfer it to the primary broth. Additional tubes of #1249 can be inoculated by transferring 0.5 ml of the primary broth using good anaerobic technique. Plate 0.1 ml of the inoculated culture onto a non-selective medium and incubate aerobically at 37°C. Incubate the anaerobic tube at 37°C.

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6. Growth should be detected in the #1249 broth within 24 to 48 hours. There should be no growth detected on the aerobic plates.

ANAEROBIC CONDITIONS:

a. To obtain a fully reduced medium, it is necessary that the medium be anoxic and that a reducing agent be added. Common reducing agents are sodium sulfide, 3% cysteine, or 1.5% sodium sulfide (stock solutions), Add 0.1 ml of reducing agent for each 10 ml of medium. For BAA-1916, cysteine worked well as a reducing agent.

b. Syringes can be made anaerobic by one of two methods. 1. Displace the dead space in the syringe with a sterile

Notes

Motile cells that are vibrioid in shape occur singly and in paris.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Desulfovibrio* sp. (ATCC BAA-1916)

References

References and other information relating to this material are available at www.atcc.org.

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