



# **Anaerobacillus arseniciselenatis (Switzer Blum et al.) Zavarzina et al.**

**700614™**

## **Description**

**Strain designation:** E1H

**Deposited As:** *Bacillus arseniciselenatis* Blum et al.

**Type strain:** Yes

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## **Storage Conditions**

**Product format:** Freeze-dried

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## **Intended Use**

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## **BSL 1**

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ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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**Certificate of Analysis**

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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**Growth Conditions****Medium:**

ATCC Medium 2100: Bacillus haloalkaliphile medium

**Temperature:** 30°C**Atmosphere:** Anaerobic**Handling Procedures**

1. Sterilize the top of the Balch tube by spraying it with 70% ethanol and then flaming the top.
2. If needed exchange the gas in the test tube for 100% N<sub>2</sub>.
3. If the medium is pink (see discussion about resazurin) add 2.0 ml of reducing

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agent (3% cysteine, stock solution) per 100 ml of medium. Let the medium sit at room temperature for 10 to 20 minutes, until the resazurin becomes colorless, before inoculating.

4. Open the freeze-dried vial according to the enclosed instructions. Take a gassed 1.0 ml syringe tipped with 22 gauge needle and withdraw 0.5 ml of medium from the Balch tube and rehydrate the freeze dried pellet. Immediately place the re-hydrated vial under a stream of sterile gas, 80% H<sub>2</sub>-20% CO<sub>2</sub> to maintain anaerobicity.

5. Using the same syringe, withdraw the cell suspension from the vial and transfer it to the Balch tube. Plate

0.1 ml of the inoculated culture onto a non-selective medium and incubate aerobically at 30°C.

6. Growth should be detected in the broth within 48 to 96 hours. No growth should be detected on the aerobic plate or broth.

**ANAEROBIC CONDITIONS:**

A. Balch tubes (available from Bellco Glass, Vineland, NJ; are specially designed for anaerobic work and use an aluminum crimp cap to hold a rubber stopper in place. Needles can easily be inserted through the stopper, and the tubes can be pressurized to 2 atm. Alternatively, serum vials may be used, or screw cap tubes with butyl rubber stoppers, in the latter case the stopper may be removed and the tube placed under a cannula system that dispenses sterile, oxygen free gas for addition of reducing agents or inoculation.

B. Resazurin is a commonly used redox indicator that is pink when the redox potential is above 50 mv, and colorless when the redox potential is below 110 mv. i.e. highly reducing. Most strict anaerobes require this low redox potential for optimum growth.

C. To obtain a fully reduced medium, it is necessary that the medium be anoxic and that a reducing agent be added. Common reducing agents are sodium sulfide, cysteine, dithiothreitol, and titanium citrate.

D. Syringes can be made anaerobic by one of two methods.

1. Displace the dead space in the syringe with a sterile

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2. Displace the dead space in the syringe with a

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**Material Citation**

If use of this material results in a scientific publication, please cite the material in the following manner: *Anaerobacillus arseniciselenatis* (Switzer Blum et al.) Zavarzina et al. (ATCC 700614)

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**References**

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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**Revision**

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**Contact Information**

ATCC

10801 University Boulevard

Manassas, VA 20110-2209

USA

US telephone: 800-638-6597

Worldwide telephone: +1-703-365-2700

Email: [tech@atcc.org](mailto:tech@atcc.org) or contact your local distributor

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