



# *Ectothiorhodospira haloalkaliphila* Imhoff and Suling

51935™

Product Sheet

## Description

**Strain designation:** BN9903 [Imhoff 51/7]

**Deposited As:** *Ectothiorhodospira haloalkaliphila* Imhoff and Suling

**Type strain:** Yes

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## Storage Conditions

**Product format:** Frozen

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## Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## Growth Conditions

### Medium:

ATCC Medium 1991: Modified halophilic *Ectothiorhodospira* medium

**Temperature:** 30°C

**Incubation:** Under a tungsten lamp

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## Handling Procedures

1. Open vial according to enclosed instructions. This organism can tolerate brief exposure to oxygen, so it may be opened under aerobic conditions. Aseptically transfer 0.5 ml of medium #1991 to the vial and rehydrate the pellet. Transfer this suspension back into a single, small tube (filled to capacity) of #1991 broth. Plate 0.1 ml of the culture on any non-selective media and incubate aerobically in the dark at 30°C. Seal the tube with a screw cap, and incubate at 30°C (room temperature)

under a tungsten lamp.

3. After four to seven days, growth is evident by turbidity. When examined microscopically, the cells appear as vibroid rods, in singles and pairs, some small chains. Once growth has been detected, the culture should be transferred to fresh broth. Subsequent growth is detected within 48 to 72 hours.

4. For best results the medium should be pre-reduced, this can best be accomplished by :

a) Steaming the medium then filling the test tube to capacity and sealing with a screw cap while the medium is still hot.

b) Adding 5 to 6 drops of cysteine (3% stock concentration) or sodium sulfide (1.5% stock concentration) to each 5 to 6 ml of medium used. After the reducing agent (cysteine or sodium sulfide) has been added fill the test tube to capacity and seal with a screw cap. Let the test tube sit at room temperature for 30 minutes before inoculating.

5. Note that once growth has been established, the culture can be transferred without pre-reducing the medium by using a 10 to 20% inoculation.

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## Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Ectothiorhodospira haloalkaliphila* Imhoff and Suling (ATCC 51935)

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## References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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