



Mycoplasmopsis hyopharyngis (Erickson et al.) Gupta et al.

51909TM

Description

Mycoplasmopsis hyopharyngis strain H3-6B-F is a bacterial type strain that was isolated from swine pharynx.

Strain designation: H3-6B-F

Deposited As: *Mycoplasma hyopharyngis* Erickson et al.

Type strain: Yes

Storage Conditions

Product format: Freeze-dried

Storage conditions: 2°C to 8°C

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 2

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories* (BMBL), U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local



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or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:

ATCC Medium 0988: Spiroplasma medium SP-4

Temperature: 37°C

Atmosphere: 95% Air, 5% CO₂

Handling Procedures

1. Follow instructions as suggested for the culturing of *Mollicutes*:

PROCEDURES FOR PROPAGATING *MOLLICUTES*:



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- a. Open the vial according to the enclosed instructions.
- b. Using a Pasteur or 1.0 ml pipette, withdraw approximately 0.5 to 1.0 ml from a tube containing 5.0 ml. Rehydrate the pellet.
- c. Aseptically transfer this aliquot back into the tube. Mix well.
- d. Make serial dilutions by transferring 0.5 mL from the original tube to a tube containing 4.5 mL. Repeat process by transferring 0.5 mL from the second to a third tube, etc. Dilutions are important, not only for titration purposes, but also to keep culture in varying stages of growth. Many strains will die out rapidly once acid or alkaline conditions are reached. It is recommended to prepare several dilutions from the initial tube as the cryoprotectant used in the freeze-drying process often inhibits growth.
- e. Use an uninoculated tube of broth to serve as a control.
- f. Plates may be inoculated to check colonial morphology. You can also spot each dilution on the surface of plate (4 or more/plate) to determine the number of colony-forming units. However, not all strains do well on solid medium.
- g. Incubate all tubes and plates under the recommended conditions and appropriate temperature. The time necessary for growth will vary from strain to strain. Growth on plates generally requires additional incubation.
- h. Depending on the medium used, growth will be indicated by increased turbidity, a color change, or both.

2. For long term storage of *Mycoplasma* species, freeze-drying or freezing is recommended. Liquid nitrogen storage is the best method. Optimally grown cells are centrifuged at 9000 rpm for 30 minutes, the supernatant poured off, and the packed cells resuspended in a smaller amount of the recommended broth. To this, add an equal amount of sterile 20% glycerol as a cryoprotectant. This suspension is aliquoted into small plastic vials and stored at -70°C or below.

Notes

Tubes are incubated aerobically. Plates must be incubated under CO₂.

This strain does not show any turbidity or color change in broth medium. To monitor growth, plates must be inoculated and observed daily for colony formation. Initial growth will take 48 to 96 hours with subsequent transfers growing in 48 hours.

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Store vials at freezer temperatures until ready to use. Additional information on this culture is available on the ATCC web site at www.atcc.org.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Mycoplasmopsis hyopharyngis* (Erickson et al.) Gupta et al. (ATCC 51909)

References

References and other information relating to this material are available at www.atcc.org.

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