



Dermamoeba minor **(Pussard et al.) Page**

50926™

Description

Strain designation: GMU-1

Deposited As: *Dermamoeba minor* (Pussard et al.) Page

Type strain: No

Storage Conditions

Product format: Frozen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always

used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:

ATCC Medium 919: Non-nutrient agar

Instructions for complete medium: 1/20th strength ATCC Medium 1405 inoculated with *Chlorella* sp. ATCC[®] 30562 as a food source (see product sheet for this strain for cultivation details).

Temperature: 25°C

Handling Procedures

Frozen ampules packed in dry ice should either be thawed immediately or stored in liquid nitrogen. If liquid nitrogen storage facilities are not available, frozen ampules may be stored at or below -70°C for approximately one week. **Do not under any circumstance store frozen ampules at refrigerator freezer temperatures (generally -20°C).** Storage of frozen material at this temperature will result in the death of the

culture.

1. To thaw a frozen ampule, place in a 35°C water bath, until thawed (2-3 min). Immerse the ampule just sufficient to cover the frozen material. Do not agitate the ampule.

2. Immediately after thawing, aseptically transfer contents to a screw-capped borosilicate test tube containing ATCC Medium 5. Incubate the tube on a 15° horizontal slant at 50-100 $\mu\text{Einsteins}/\text{m}^2/\text{s}$ irradiance at 25°C with the cap loosened one half turn. Maintain under a 14/10 h light-dark photoperiod and subculture every 14-21 days.

Culture maintenance:

1. For a slant culture, transfer cells with an inoculating loop to a tube of fresh agar medium from a growing culture at or near peak density. For a broth culture, inoculate a tube of fresh broth medium with 0.1 ml from a growing culture at or near peak density.

2. Incubate at 50-100 $\mu\text{Einsteins}/\text{m}^2/\text{s}$ irradiance at 25° C with the cap loosened one half turn. Maintain under a 14/10 h light-dark photoperiod and subculture every 14-21 days.

Cryopreservation:

1. Harvest cells from a culture which is at or near peak density by adding 3.0 ml fresh ATCC medium 5 broth to the slant and washing cells into suspension.

2. Adjust the concentration of cells to 2×10^6 - 2×10^7 / ml with fresh broth medium, then dilute to half this concentration by adding an equal amount of a 10% (v/v) sterile methanol solution in fresh ATCC medium 5 broth.

3. Dispense in 0.5 ml aliquots into 1.0 - 2.0 ml sterile plastic screw-capped cryules (special plastic vials for cryopreservation). The time from the mixing of the cell preparation and methanol stock solution to the beginning of the freezing process should be no less than 5 min and no greater than 15 min.

4. Place vials in a controlled rate freezing unit. From room temperature cool at -1°C/min to -40°C. If freezing unit can compensate for the heat of fusion, maintain rate at -1 C/min through heat of fusion. At -40°C plunge ampules into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the

apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately -1°C/min.)

5. The frozen preparations should be stored in either the vapor or liquid phase of a nitrogen refrigerator. Frozen preparations stored below -130°C are stable indefinitely. Those stored at temperatures above -130°C are progressively less stable as the storage temperature is elevated. Vials can be stored between -80 and -70°C for no longer than one week.

6. To establish a culture from the frozen state place an ampule in a water bath set at 35°C. Immerse the vial to a level just above the surface of the frozen material. Do not agitate the vial.

7. Immediately after thawing, do not leave in the water bath, aseptically remove the contents of the ampule and add to a centrifuge tube containing 5 ml of ATCC medium 5 without agar. Centrifuge at 300 x g for 5 min.

10. Remove most of the supernatant (=methanol, which can inhibit growth) and then resuspend the pellet. Transfer the culture to a fresh tube of ATCC medium 5 and incubate on a 15° horizontal slant at 50-100 μ Einsteins/m²/s irradiance at 25°C with the cap loosened one half turn. Maintain under a 14/10 h light-dark photoperiod and subculture every 14-21 days.

Notes

Additional information on this culture is available on the ATCC web site at www.atcc.org.

While every effort is made to insure authenticity and reliability of strains on deposit, ATCC is not liable for damages arising from the misidentification or misrepresentation of cultures.

ATCC recommends that individuals contemplating commercial use of any culture first contact the originating investigator to negotiate an agreement. Third party distribution of this culture is discouraged, since this practice has resulted in the unintentional spreading of contaminated cultures.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Dermamoeba minor* (Pussard et al.) Page (ATCC 50926)

References

References and other information relating to this material are available at www.atcc.org.

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