Helicobacter bilis Fox et al.

43879™

Description

Strain designation: SLH-38264
Deposited As: *Flexispira rappini* Bryner et al.
Type strain: No

Storage Conditions

Product format: Freeze-dried

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 2

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always

used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:
ATCC Medium 18: Trypticase Soy Agar/Broth
ATCC Medium 260: Trypticase soy agar/broth with defibrinated sheep blood
Temperature: 37°C
Atmosphere: Microaerophilic: 3-5% O_2, 10% CO_2

Handling Procedures

1. Open vial according to enclosed instructions. Rehydrate contents of vial with 0.5 mL of #18 broth.
2. To obtain a biphasic culture, add 0.4 mL of the suspension to a #260 slant. Add remaining 0.1 mL of the suspension to a #260 plate and streak for isolation.
3. Incubate at 37°C under microaerophilic conditions using an anaerobe jar with an active catalyst and a microaerophilic gas generator pack, or other

acceptable method, to obtain microaerophilic conditions. Incubate slant with cap loose.

4. Within three to five days of incubation, good growth should be obtained in the broth pool at the bottom of the slant. Secondary subcultures to fresh biphasic slants require shorter incubation times, 24 to 48 hours. Additional incubation may be required for colonies to appear on the plate. Further subcultures can be made using broth pool as the inoculum source.

Notes

This is a slow growing organism that requires moist conditions for best growth. Growth at the broth/agar interface of the biphasic slant should occur within three to five days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy. The organism is a medium size, fusiform shaped, motile bacillus.

Individual colonies will not be observed. Growth appears as a film on the surface of fresh, moist plates, extending out from original inoculation site to fill entire plate, usually within 48 to 72 hours.

The cells do not Gram stain well using traditional procedures. For best results, use a basic fuchsin counterstain in place of the safranin.

Once good growth is obtained, transfer or freeze the culture. Adding an equal amount of 20% sterile glycerol to pooled broth from several biphasic slants followed by freezing in liquid nitrogen or "ultra-low temperature" freezer is recommended.

Additional information on this culture is available on the ATCC® web site at www.atcc.org.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Helicobacter bilis* Fox et al. (ATCC 43879)

References

References and other information relating to this material are available at www.atcc.org.

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