



# ***Campylobacter jejuni* subsp. *jejuni* (Jones et al.) Steele and Owen**

**43445™**

## **Description**

**Strain designation:** MK52

**Deposited As:** *Campylobacter jejuni* subsp. *jejuni* (Jones et al.) Veron and Chatelain

**Type strain:** No

**Serotype:** O:18

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## **Storage Conditions**

**Product format:** Freeze-dried

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## **Intended Use**

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## **BSL 2**

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

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ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## **Certificate of Analysis**

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## **Growth Conditions**

### **Medium:**

ATCC Medium 1116: Brucella broth with 0.16% agar

**Temperature:** 37°C

**Atmosphere:** Microaerophilic

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## **Handling Procedures**

1. Open vial according to enclosed instructions.
2. Using a single tube of #1115, #177, or #18 broth (5 to 6 ml), withdraw approximately 0.5 to 1.0 ml with a Pasteur or 1.0 ml pipette. Rehydrate the entire pellet.
3. Aseptically transfer this aliquot back into the broth tube. Mix well.

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**4. Use several drops of the suspension to inoculate a #260 agar slant and/or plate.**

5. Or, to obtain a biphasic culture, add 0.5 ml of the suspension to a #260 agar slant (see notes)

6. Incubate tubes and plate at 37°C, under microaerophilic conditions, for 24 to 48 hours. Use an anaerobe jar with an active catalyst and a microaerophilic gas generator pack, or other acceptable method. Incubate slant with cap loose.

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## **Notes**

Colonies on #260 agar plates are entire, circular, smooth, glistening, and low convex.

This is an organism that requires moist conditions for best growth. A biphasic culture gives the most rapid growth. Growth at the broth/agar interface of the biphasic slant should occur within one to two days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy. The organism is a short, thin motile rod. Motility is usually observed only in young cultures.

Growth on agar takes longer than with the biphasic culture. Once good growth is present, these organisms tend to lose viability, especially if exposed to air for lengthy periods.

Additional information on this culture is available on the ATCC® web site at [www.atcc.org](http://www.atcc.org).

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## **Material Citation**

If use of this material results in a scientific publication, please cite the material in the following manner: *Campylobacter jejuni* subsp. *jejuni* (Jones et al.) Steele and Owen (ATCC 43445)

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## **References**

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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## **Revision**

This information on this document was last updated on 2025-08-16

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## **Contact Information**

ATCC

10801 University Boulevard

***Campylobacter jejuni* subsp. *jejuni* (Jones et al.) Steele and Owen**

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Manassas, VA 20110-2209

USA

US telephone: 800-638-6597

Worldwide telephone: +1-703-365-2700

Email: [tech@atcc.org](mailto:tech@atcc.org) or contact your local distributor

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