



# ***Ureaplasma parvum*** **Robertson et al.**

**33697<sup>TM</sup>**

## **Description**

*Ureaplasma parvum* strain U26 was isolated from a pregnant woman. This strain requires an additional 5-10% horse serum added to the broth for growth.

**Strain designation:** U26

**Deposited As:** *Ureaplasma urealyticum* Shepard et al.

**Type strain:** No

**Serotype:** 14

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## **Storage Conditions**

**Product format:** Freeze-dried

**Storage conditions:** 2°C to 8°C

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## **Intended Use**

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## **BSL 2**

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories* (BMBL), U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local

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or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## **Certificate of Analysis**

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## **Growth Conditions**

### **Medium:**

ATCC Medium 1092: B-broth medium for *Ureaplasma*

**Temperature:** 37°C

**Atmosphere:** Broth: Aerobic; Plates: Anaerobic

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## **Handling Procedures**

1. Follow instructions as suggested for the culturing of *Mollicutes*:

PROCEDURES FOR PROPAGATING *MOLЛИCUTES*:

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- a. Open the vial according to the enclosed instructions.
- b. Using a Pasteur or 1.0 mL pipette, withdraw approximately 0.5 to 1.0 mL form a tube containing 2.5 mL of #1092 broth. Rehydrate entire pellet.
- c. Aseptically transfer this aliquot back into the tube. Mix well.
- d. Make serial dilutions by transferring 0.25 mL from the original tube to a test tube containing 2.25 mL. Repeat process by transferring 0.25 mL from the second to third tube, etc.
- e. Use an uninoculated tube of broth to serve as a control.
- f. Plates may be inoculated to check colony morphology. You can also spot each dilution on the surface of plate to determine the number of colony-forming units. However, not all strains do well on solid medium.
- g. Incubate all tubes and plates under the recommended conditions and appropriate temperature (see step 2). The time necessary for growth will vary from strain to strain. Growth in broth for this strain will generally be observed at 16 to 48 hours. Growth on plates generally requires 7 to 10 days.
- h. Depending on the medium used, growth will be indicated by increased turbidity, a color change, or both (see notes section).
- i. Inoculate a Trypticase Soy Agar with 5% Defibrinated Sheep Blood plate with 0.1 mL to check for contamination. Incubate plate at 37°C. A second plate may be incubated in the jar with the growth plates. No growth should occur on Trypticase Soy Agar with 5% Defibrinated Sheep Blood.

2. Tubes may be incubated aerobically, but plates require increased CO<sub>2</sub> (10% or higher) for growth. This is ideally achieved using an anaerobe jar with gas generating sachets. The incubation temperature is 37°C.

**Notes**

This strain requires an additional 5-10% horse serum (Thermo-Fisher, 16050-122) added to the broth for growth.

*Ureaplasma* strains grow very rapidly. The indicator in the first tube will change color to a dark green within hours. The culture should be transferred when the broth turns a light to medium green. If it is allowed to turn dark green or blue-green, it has become too alkaline. It is especially important to make serial dilutions of this strain, for when alkaline conditions are reached (as indicated by the color change), the

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culture will rapidly die unless refrigerated immediately (+4°C) or stored frozen at -60°C. Refrigerated broth cultures may remain viable for periods up to 4 days. No visible turbidity will be seen. The color change is the only indication of growth in broth. Therefore, transfer, freeze, or lyophilize the culture as soon as possible. There should be no growth on GM agar (Genital Mycoplasma) medium. Broth is the best method for propagation.

Additional information on this culture is available on the ATCC® web site at [www.atcc.org](http://www.atcc.org).

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### **Material Citation**

If use of this material results in a scientific publication, please cite the material in the following manner: *Ureaplasma parvum* Robertson et al. (ATCC 33697)

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### **References**

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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