



Trypanosoma lewisi (Kent) Laveran and Mesnil

30147™

Description

Strain designation: New Orleans-67

Deposited As: *Trypanosoma lewisi* (Kent) Laveran and Mesnil

Type strain: No

Storage Conditions

Product format: Frozen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:

ATCC Medium 1029: LIT medium

Instructions for complete medium: ATCC Medium 1029 supplemented with an additional 5-10 µg/mL hemin.

Note: This strain grows poorly without the additional hemin in the growth medium

Temperature: 25°C

Culture system: Axenic

Handling Procedures

Frozen ampules packed in dry ice should either be thawed immediately or stored in liquid nitrogen. If liquid nitrogen storage facilities are not available, frozen ampules

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may be stored at or below -70°C for approximately one week. **Do not under any circumstance store frozen ampules at refrigerator freezer temperatures (generally -20°C).** Storage of frozen material at this temperature will result in the death of the culture.

1. To thaw a frozen ampule, place in a 35°C water bath until thawed (2-3 min). Immerse the ampule just sufficiently to cover the frozen material. Do not agitate the ampule.
2. Immediately after thawing, aseptically transfer contents to a T-25 tissue culture flask containing 10 mL ATCC medium 1029 supplemented with an additional 5-10 µg/mL hemin. Incubate at 25°C with the cap screwed on tightly.

Culture maintenance:

1. Vigorously agitate a culture at or near peak density and aseptically transfer 0.1-0.2 mL to a fresh flask containing 10 mL ATCC medium 1029 supplemented with an additional 5-10 µg/mL hemin.
2. Incubate at 25°C with the cap screwed on tightly.
3. Subculture every 10-14 days as described in step 1. The transfer interval will depend on the quantity of the inoculum and the quality of the medium. This should be empirically determined by examining the culture on a daily basis until the growth cycle has stabilized.

Cryopreservation:

1. Harvest cells from a culture which is at or near peak density by centrifugation at 1,300 g for 5 min.
2. Adjust concentration of cells to 2×10^7 /mL in fresh medium.
3. While cells are centrifuging prepare a 10% (v/v) solution of sterile DMSO in fresh medium (broth). The DMSO solution when first prepared will warm up due to chemical heat. The solution should be allowed to return to room temperature prior to use.
4. Mix the cell preparation and the DMSO solution in equal portions. The final concentration will be 10^7 cells/mL and 5% (v/v) DMSO. The time from the mixing of the cell preparation and DMSO stock solution before the freezing process is begun should be no more than 15 min.
5. Dispense in 0.5 mL aliquots into 1.0 - 2.0 mL sterile plastic screw-capped cryules (special plastic vials for cryopreservation).
6. Place the ampules in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately -1°C/min.)
7. Store in either the vapor or liquid phase of a nitrogen refrigerator.

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8. To thaw a frozen ampule, place it in a 35°C water bath such that the lip of the ampule remains above the water line. Thawing time is approximately 2 to 3 minutes. Do not agitate the ampule. Do not leave ampule in water bath after thawed.
 9. Immediately after thawing, do not leave in the water bath, aseptically transfer the contents of the ampule into a fresh flask containing 10 mL ATCC medium 1029 supplemented with an additional 5-10 µg/mL hemin.
 10. Incubate at 25°C with the cap screwed on tightly.
 11. Maintain as described above.
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Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Trypanosoma lewisi* (Kent) Laveran and Mesnil (ATCC 30147)

References

References and other information relating to this material are available at www.atcc.org.

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