



Trypanosoma brucei *gambiense* Dutton

30025™

Product Sheet

Description

Strain designation: Wellcome Ts

Deposited As: *Trypanosoma gambiense* Dutton

Type strain: No

Storage Conditions

Product format: Frozen

Storage conditions: -80°C or colder for 1 week, vapor phase of liquid nitrogen for long-term storage

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 2

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Host:

in vivo cultivation, laboratory rat

Handling Procedures

Storage and Culture Initiation

Frozen ampules packed in dry ice should either be thawed immediately or stored in liquid nitrogen. If liquid nitrogen storage facilities are not available, frozen ampoules may be stored at or below -70°C for approximately one week. **Do not under any circumstance store frozen ampules at refrigerator freezer temperatures (generally -20°C).** Storage of frozen material at this temperature will result in the death of the culture.

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1. To thaw a frozen ampule, place it in a 35°C water bath such that the lip of the ampule remains above the water line. Thawing time is approximately 2 to 3 minutes. Do not agitate the ampule. Do not leave ampule in water bath after it is thawed.
2. Immediately after thawing, aseptically remove the entire contents of the ampule with a syringe and inject intraperitoneally into an adult rat known to be free of blood parasites. Follow the protocol for maintenance *in vivo*. The course of infection may be longer or shorter than usual depending on percent recovery of the parasite from the frozen state.

Culture maintenance:

1. Monitor the course of infection by drawing a drop of blood from the tail of an infected animal, mixing with 1.0 mL of buffered saline (0.8% saline buffered to pH 7), and observing microscopically. If one or more parasites are seen, follow the procedures in step 2.
2. To infect additional rats, draw a drop of blood from the tail vein of an infected animal into a syringe containing 0.5 mL of sterile buffered anticoagulant, mix, and inoculate 0.1-0.2 mL of the suspension intraperitoneally into an adult rat known to be free of blood parasites.
3. Parasites for inoculation should be collected at or near peak parasitemia (usually within 4-8 days of infection if large inocula are given). Peak parasitemia from the thawed material may be slightly sooner or later. Host rats should be approximately 4-6 months old.

Reagents for cryopreservation: Tyrode's Salt Solution

NaCl, 8.00 g

KCl, 0.20 g

CaCl₂, 0.20 gMgCl₂ · H₂O, 0.05 gNaH₂PO₄ · H₂O 1.00 gNaHCO₃ · H₂O, 1.00 g

Glucose, 1.00 g

Glass distilled H₂O to 1.00 L

Add ingredients in the sequence listed. Filter-sterilize.

Harvest and Preservation

1. **Cryopreservation:**
2. Harvest the parasites according to the protocol for maintenance *in vivo*.
3. Spin the cell suspension at approximately 50 x g for 3 min, to remove any

cellular debris.

4. Transfer the supernatant to a new 15 mL plastic centrifuge tube. Centrifuge at 1300 x g for 10 min.
5. Pool the cell pellets and adjust the concentration to $2.0 - 4.0 \times 10^7$ cells/mL with a fresh solution of Tyrode's Salt Solution. *If the concentration is too low centrifuge at 1300 x g for 10 min and resuspend in the volume of Tyrode's Salt Solution required to yield the desired concentration.
6. Mix the cell preparation and 10% (v/v) DMSO in equal portions. The final concentration will be $1.0 - 2.0 \times 10^7$ cells/mL and 5% DMSO. The time from the mixing of the cell preparation and cryoprotective solution to the start of the freezing process should be no less than 15 min. and no more than 30 min.
7. Dispense in 0.5 mL aliquots to 1.0-2.0 mL sterile plastic screw-capped cryules (special plastic vials for cryopreservation).
8. Place the vials in a controlled rate freezing unit. From room temperature cool at $-1^\circ\text{C}/\text{min}$ to -40°C . If the freezing unit can compensate for the heat of fusion, maintain rate at $-1^\circ\text{C}/\text{min}$ through the heat of fusion. At -40°C plunge into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately $-1^\circ\text{C}/\text{min}$.)
9. Store in either the vapor or liquid phase of a nitrogen refrigerator.
10. To thaw a frozen ampule, place it in a 35°C water bath such that the lip of the ampule remains above the water line. Thawing time is approximately 2 to 3 minutes. Do not agitate the ampule. Do not leave ampule in water bath after thawed.
11. Immediately after thawing, aseptically remove the contents of the ampule with a syringe and inoculate an uninfected rat. Follow the protocol for maintenance *in vivo*.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Trypanosoma brucei gambiense* Dutton (ATCC 30025)

References

References and other information relating to this material are available at www.atcc.org.

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