



Acholeplasma oculi Al-Aubaidi et al.

27350™

Description

Strain designation: [19-L]

Deposited As: *Acholeplasma oculi* Al-Aubaidi et al.

Type strain: Yes

Storage Conditions

Product format: Freeze-dried

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 2

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always

used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:

ATCC Medium 639: Heart infusion agar or broth, pH 7.5, containing 20% (v/v) inactivated- and acid-adjusted porcine serum and 10% (v/v) yeast extract

Temperature: 37°C

Atmosphere: 95% Air, 5% CO₂

Handling Procedures

1. Follow instructions as suggested for the culturing of

Mollicutes:

PROCEDURES FOR PROPAGATING *MOLLICUTES*:

a) Open the vial according to the enclosed instructions.

b) Using a Pasteur or 1.0 ml pipette, withdraw

approximately 0.5 to 1.0 ml from a tube containing

5.0 ml. Rehydrate the entire pellet.

c) Aseptically transfer this aliquot back into the tube. Mix well.

d) Make serial dilutions by transferring 0.5 ml from the

original tube to a tube containing 4.5 ml. Repeat process by transferring 0.5 ml from the second to a third tube, etc. Dilutions are important, not only for titration purposes, but also to keep culture in varying stages of growth. Many strains will die out rapidly once acid or alkaline conditions are reached. It is recommended to prepare several dilutions from the initial tube as the cryoprotectant used in the freeze-drying process often inhibits growth.

e) Use an uninoculated tube of broth to serve as a control.

f) Plates may be inoculated to check colonial morphology. You can also spot each dilution on the surface of plate (4 or more/plate) to determine the number of colony-forming units. However, not all strains do well on solid medium.

g) Incubate all tubes and plates under the recommended conditions and appropriate temperature. The time necessary for growth will vary from strain to strain. Growth on plates generally requires additional incubation.

h) Depending on the medium used, growth will be indicated by increased turbidity, a color change, or both.

2. Tubes are incubated aerobically, plates are incubated under 5% CO₂ or in a *candle jar*. The incubation temperature is 37°C. Growth will initiate in the first few tubes within 24 hours.

3. For long term storage of *Acholeplasma* sp., freeze-drying or freezing is recommended. Liquid nitrogen storage is the best method. Optimally grown cells are centrifuged at 9000 rpm for 30 minutes, the supernatant poured off, and the packed cells resuspended in a smaller amount of #639 broth. To this, add an equal amount of sterile 20% glycerol as a cryoprotectant. This suspension is aliquoted into small plastic vials and stored at -70°C or below.

Notes

Store vials at freezer temperatures until ready to use.

We have found that using a *candle jar* for CO₂ conditions works better for those strains whose medium has an indicator present. CO₂ incubators may lower the pH of the medium enough to cause a color change. This change may make it difficult to observe growth with those strains that show little turbidity.

Additional information on this culture is available on the ATCC web site at www.atcc.org.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Acholeplasma oculi* Al-Aubaidi et al. (ATCC 27350)

References

References and other information relating to this material are available at www.atcc.org.

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