Desulfovibrio salexigens Postgate and Campbell

14944™

Description

Strain designation: NCIB 8365 [Louisiana 43.11]
Deposited As: *Desulfovibrio salexigens* Postgate and Campbell
Type strain: No

Storage Conditions

Product format: Freeze-dried
Storage conditions: 2°C to 8°C

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:
ATCC Medium 1250: Modified Barr's Medium for sulfate reducers with 2.5% NaCl
Temperature: 37°C
Atmosphere: 80% N_2, 20% CO_2
Incubation: With shaking

Handling Procedures

1. Open vial according to enclosed instructions.
2. Perform all steps under anaerobic conditions. (*see below*). Exchange the gas in the head space for a fresh anaerobic gas, either 80% N_2 20% CO_2 or 100% N_2. To insure that the media is anaerobic add 0.1 mL 1.5% sodium sulfide (stock concentration) for each 5 to 10 mL of medium.

3. Using an anaerobic 1 mL syringe (see below) aseptically transfer 0.5 mL of ATCC Medium #1250 to the vial and rehydrate the entire freeze-dried pellet. Transfer the entire suspension back into the primary tube. The primary tube should not contain more than 8 to 10 mL of #1250 broth. Secondary tube(s) can be inoculated with 0.5 mL of the primary broth. Inoculate a plate of non-selective medium with 0.1 of the culture.
4. Incubate the tubes anaerobically at 37°C with gentle shaking. Incubate the plate(s) aerobically as a purity check.
5. After two or three days, growth should be evident as indicated by turbidity throughout the broth. Once growth has been established, the culture should be transferred to fresh broth every 24 to 48 hours.
6. This culture is sensitive to oxygen, therefore steps should be taken to avoid exposure to oxygen. When the culture exhibits good growth it will remain viable for up to 1 week if stored at 4°C under anaerobic condition.

ANAEROBIC CONDITIONS:

A. Balch tubes (available from Bellco Glass, Vineland, NJ; are specially designed for anaerobic work and use an aluminum crimp cap to hold a rubber stopper in place. Needles can easily be inserted through the stopper. Alternatively, serum vials may be used, or screw cap tubes with butyl rubber stoppers, in the latter case the stopper may be removed and the tube placed under a cannula system that dispenses sterile, oxygen free gas for the addition of reducing agents or inoculation.
B. To obtain a fully reduced medium, it is necessary that the medium be anoxic and that a reducing agent be added. Common reducing agents are sodium sulfide, cysteine, dithiothreitol, titanium citrate and Co-enzyme M (see D). If component IV is added to the medium sodium sulfide, dithiothreitol and titanium citrate will cause the ferrous ammonium sulfate to precipitate even without growth.
C. We suggest adding the reducing agent to the medium at least one hour before the medium is to be inoculated.
D. Syringes can be made anaerobic by one of two methods.
    1. Displace the dead space in the syringe with a sterile oxygen-free gas.
    2. Displace the dead space in the syringe with a reducing agent.

Notes

Growth should be detected within 24 to 48 hours as indicated by turbidity throughout the broth. Turbidity may not be evident at 24 hours in the secondary growth tubes, but viable cells can be observed microscopically.

The cells typically appear as comma-shaped rods that are motile.

Once growth has been established the culture should be transferred every 24 hours when maintained at 37°C. The culture can be maintained at 4°C for up to 1 week.

Additional information on this culture is available on the ATCC® web site at www.atcc.org.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Desulfovibrio salexigens* Postgate and Campbell (ATCC 14944)

References

References and other information relating to this material are available at www.atcc.org.

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Revision

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